

Review

Seaweed-Based Molecules and Their Potential Biological Activities: An Eco-Sustainable Cosmetics

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Abstract: Amongst the countless marine organisms, seaweeds are considered as one of the richest sources of biologically active ingredients having powerful biological activities. Seaweeds or marine macroalgae are macroscopic multicellular eukaryotic photosynthetic organisms and have the potential to produce a large number of valuable compounds, such as proteins, carbohydrates, fatty acids, amino acids, phenolic compounds, pigments, etc. Since it is a prominent source of bioactive constituents, it finds diversified industrial applications viz food and dairy, pharmaceuticals, medicinal, cosmeceutical, nutraceutical, etc. Moreover, seaweed-based cosmetic products are risen up in their demands by the consumers, as they see them as a promising alternative to synthetic cosmetics. Normally it contains purified biologically active compounds or extracts with several compounds. Several seaweed ingredients that are useful in cosmeceuticals are known to be effective alternatives with significant benefits. Many seaweeds' species demonstrated skin beneficial activities, such as antioxidant, anti-melanogenesis, antiaging, photoprotection, anti-wrinkle, moisturizer, antioxidant, anti-inflammatory, anticancer and antioxidant properties, as well as certain antimicrobial activities, such as antibacterial, antifungal and antiviral activities. This review presents applications of bioactive molecules derived from marine algae as a potential substitute for its current applications in the cosmetic industry. The biological activities of carbohydrates, proteins, phenolic compounds and pigments are discussed as safe sources of ingredients for the consumer and cosmetic industry.

Keywords: cosmeceuticals; seaweeds; skin cosmetics; marine macroalgae; biological activities

1. Introduction

Cosmeceuticals are defined by cosmetic producers as products to improve or alter the skin functions and appearance, causing skin benefits [1]. The term "cosmeceutical" refers to products that can combine both cosmetic and pharmaceutical uses to improve skin characteristics, such as the appearance, structure, and functions of the skin [2,3]. This cosmeceutical sector is highly innovative and always looking for principally active molecules that serve better characteristics to open up many possibilities [4]. The cosmetic sector continues to develop in many developing countries that started by the global beauty market [5].

Based on this encouraging future, the production of cosmetics formulations without any side effects is practiced to satisfy the customers [6,7]. The ever-expanding market of the cosmetic industry has led to the use of synthetic or chemical compounds for economic benefits [8]. These harmful components interact with the skin layers and produce toxicological effects on the living body [9,10]. There are many synthetic chemicals occupying space in cosmetic preparation, such as BHA (Butylated hydroxyanisole), BHT (Butylated hydroxytoluene), Coal tar dyes, diethanolamine (DEA), dibutyl phthalate (DBP), parabens, perfume, polyethylene glycol (PEG), petrolatum, siloxane, and heavy metals [11–19]. These components accumulate in the skin layers and cause many dermatological conditions, such as dermatitis, cancers, skin rashes, multiple stretch marks, yellowish-brown coloration, etc. [20,21]. Because of these harmful side effects, the use of cosmetics products has become a serious public health problem. This type of use with prolonged exposure on skin accumulates and causes harmful effects, such as an allergic reaction, irritation, and exfoliation [22]. Hence, the goal is to reduce and remove these undesired effects by using natural resources as an alternative in cosmetic formulations to meet consumer demands. Natural resources offer many more advantages, as they are environmentally friendly, are less toxic, are non-carcinogenic, are easily accessible, have lesser side effects, and are economically beneficial [23–25]. These include terrestrial plants, animals, and heterogeneous groups in the oceans, etc., with their richness as a source of biologically active principles [26]. A variety of marine organisms, such as fishes, seabirds, reptiles, marine mammals, seaweeds, and many other sources, have been examined by researchers to identify and extract various biologically active constituents, and they have found that all resources are an exceptional reservoir for potential ingredients [27,28]. Among diversified marine organisms, seaweeds are utilized as one of the most significant sources with wide potentials, alternatively [29]. Seaweeds are a novel source of potentially active compounds (proteins–lectins, phycobiliproteins, peptides, amino acids, polyphenols, and polysaccharides) to be exploited in human health benefits, such as antiviral, anticancer, anticoagulant, anti-obesity, and diabetes modulator [30]. Shannon and Abu-Ghannam [31], suggested seaweed as nutraceuticals or functional foods with dietary benefits beyond their fundamental macronutrients, highlighting their significant effect on obesity and dietary related disease. This study also suggested recent developments of seaweed applications for human health from epidemiological and functional food perspectives. Brownlee et al. [32] reported the importance of alginate (algal polysaccharide) as a dietary supplement for maintenance of normal health. They also studied fiber-like activities, particularly its effects on intestinal absorption and the colon. Along with health benefits, alginate has several roles in cosmetics to help retain moisture and act as an emulsion stabilizer, bonding agent, and facial mask (filmogen) to hydrate, soothe, and soften skin. Seaweeds are similarly known as marine algae, a term encompassing macroscopic multicellular, benthic, non-flowering eukaryotic photosynthetic organisms [33–37]. Marine algae are found in diversified habitats such, as in tidal or sub-tidal regions, or shallow coastal water of the sea. They are also found attached with substrata, such as other aquatic plants, rocks, dead corals, pebbles, shells, and sand particles [38–40]. Green and red algae belong to the Plantae kingdom, whereas brown algae belong to the Chromista kingdom [41,42]. It is classified mainly in three categories based on the presence of photosynthetic pigments, such as red algae, green algae, and brown algae in Rhodophyta, Chlorophyta, and Ochrophyta phylum (Phaeophyceae class), respectively [43,44]. Many researchers suggested that marine seaweeds are effective natural alternatives to synthetic chemicals by showing many skin benefits, such as moisture retention, cell renewal activation, cell metabolism, regulation of sebaceous secretion and tissue drainage, promoting blood circulation, and increasing skin resistance [45–47]. There are many seaweed-based cosmetic products commercially available, such as Voya Get Glowing Illuminating Clay Mask (*Himanthalia elongata*), OSEA Ocean Cleansing Mudd (*Fucus vesiculosus*), Biossance Squalane + Probiotic Gel Moisturizer (*Chondrus crispus*), Repêchage Vita Cura B3 Serum Complex (*Laminaria digitata*), and Ayla Sea Soak (*Macrocystis pyrifera*). Some other products, such as True Botanicals Clear Pure Radiance Oil,

Skinceuticals Daily Moisture, Mario Badescu Seaweed Night Cream, and Dr. Dennis Gross Skincare Hyaluronic Marine Oil-Free Moisture Cushion, etc., also occupy the cosmetic market. In addition, Heo et al. [26] reported that seaweed is a promising and focused group of novel biochemically active principal molecules and nowadays appreciated in the developments of new biotechnological or cosmeceutical purposes.

The chemically diversified nature and unique potential of seaweeds are the reason why they have been the focus of interest for the past few years in various cosmetic applications. Seaweed-based protein, polysaccharides, phenolic compounds, and pigment profiles present cosmetic and cosmeceutical potential. This review study gives an overall view of an exploitation of seaweed for cosmetic beneficial activities. Mainly, the role of polysaccharide, protein, phenolic compounds, and pigments in different skin cosmetic beneficial activities are discussed.

1.1. Cosmetic Potential of the Seaweed Compounds

Marine macroalgae produce both primary metabolites, including proteins, amino acids, polysaccharides, fatty acids, etc., and secondary metabolites, such as phenolic compounds, pigments, sterols, vitamins, and other bioactive components [48–54]. Moreover, various types of biological activities expressed by different phycocompounds, such as blood coagulation system, antilipidemic activity, immunomodulating effect, antiviral activity, anticancer activity, antimicrobial activity, antioxidant activity, and other significant activities [55]. Especially in the area of cosmetics, many scientists reported skin beneficial activities, such as antiaging, anti-wrinkle, anti-cellulite, antioxidant, moisturizing, whitening, and photoprotection [56–63]. Sun and Chavan, [64] studied *Fucus vesiculosus* extract to reduce the appearance of dark circles on the skin area by enhancing the expression of hemoxygenase-1. By removing heme catabolites, it eliminates the heme production on skin. Hagino and Saito [65] reported some algae species and derived compounds for UV protection benefits, skin moisturization, and inhibition of melanin synthesis. Leyton et al. [66] identified phlorotannins, phloroeckol, and phloroglucinol in the extract of brown macroalgae *Macrocystis pyrifera*. They also reported good antidiabetic and antioxidant activity of phlorotannins, which can prevent skin aging.

Additionally, Yu and Gu [67] reported the role of algae-derived polysaccharides in the formation of protective membrane to prevent water evaporation in skin. Likewise, Sulfated polysaccharide from the red algae appear to be an excellent candidate to substitute hyaluronic acid as a bio lubricant and antioxidant [68]. Low-molecular-weight polysaccharides derived from red algae *Pyropia yezoensis* had skin beneficial functions, such as antioxidant, anti-inflammation, photoaging protection, etc. [69]. In the case of pigment, red carotenoid pigment, astaxanthin, scavenges free radicals and blocks proinflammatory cytokine production [70]. Moreover, Thomas and Kim [71] isolated fucoxanthin from *Laminaria japonica*; it is reported to inhibit tyrosinase activity and melanogenesis in UVB-irradiated mice.

1.2. Polysaccharides

The polysaccharides are the most significant and beneficial compounds present in macroalgae and characterized for their biological skin beneficial activity. Seaweeds are well-known for many different types of a polysaccharides, such as chitin, fucoidans, agar, carrageenan, alginates, ulvans, terpenoids, and tocopherol [72–77]. In skin cosmeceuticals, marine algae have exhibited activities such as anti-melanogenesis, antioxidant, anti-skin-aging, anti-inflammation, anti-atopic dermatitis, anti-skin-cancer, and repair of UV-induced damage [6,78–83]. A significant amount of carbohydrates is found in many macroalgal species, such as *Kappaphycus alvarezii* (formerly *Eucheuma cottonii*) (Rhodophyta), *Sargassum polycystum* (Phaeophyceae), *Padina boryana* (formerly *Padina tenuis*) (Phaeophyceae), *Fucus vesiculosus* (Phaeophyceae), *Porphyra umbilicalis* (Rhodophyta), etc. Moreover, polysaccharides have a wide variety of applications, such as photoprotection, moisturizer, wound-healing agents, thickening agents, emulsifiers, and preservatives [26,84–90]. Nowadays, the makers

of skincare products are focused on compounds to control or regulate potential tyrosinase inhibition, inhibition of collagenase, elastase, reduction of matrix metalloproteinase (MMP) activity, reduce reactive oxygen species (ROS), and antioxidant activity [91–96]. Table 1 exhibits different potential skin beneficial effects of seaweeds' polysaccharides. Of particular interest to the cosmeceutical utility, Fujimura et al. [28] explained the applications of purified fucoidan extracts of *Fucus vesiculosus* into creams and lotion, providing antiaging and anti-wrinkle benefits. They also reported collagenase expression, anti-inflammatory activity, and inhibition of matrix enzymes against hyaluronidase, heparinase, tyrosine kinase, and phospholipase A2. Holtkamp et al. [97] also reported the usefulness of fucoidan, particularly in skin protecting, antioxidants, antiaging, antiviral, anti-inflammatory, antitumor, and anticoagulant properties, by epidemiological and experimental studies. Fujimura et al. [98] illustrated a significant reduction in skin thickness, together with improvements in elasticity, of gel formulation with 1% *Fucus* extract. Additionally, Teixeira and Hellewell [99] revealed the use of fucoidan as a topical anti-inflammatory for cosmetic after-sun damage, allergic-condition-soothing products, or specially postsurgical formulations. Polysaccharides have also been widely shown to exhibit antioxidant, antiviral, anticoagulant, and antitumor properties in commercial products. For instance, lipid soluble fraction of an edible red alga *Gelidium amansii* to induced apoptosis of cancer cells in vitro [100]. Like fucoidan, carrageenan produces a range of textures for creams, lotions, sticks, sprays, and foams formulation [101]. It has a range of biological properties and is applicable in various pharmaceutical properties, such as antitumor, immunomodulation, anti-hyperlipidemic, and anticoagulant activities [102]. Another phenolic compound are alginates found in the cell wall of brown algae (Phaeophyceae), mainly *Laminaria* species (*Laminaria hyperboreun*, *Laminaria digitata*, *Laminaria japonica*) and also *Macrocystis pyrifera*, *Ascophyllum nodosum*, *Ecklonia maxima*, *Lessonia nigrescens*, *Ascophyllum nodosum*, *Durvillea antarctica*, and *Sargassum* sp. [98,103]. Podkorytova et al. [104] also employed alginates in cosmetics for face-mask and body-wash ingredients due to their benefits to the skin structure and function. Specifically, alginates are highly efficient when used to solidify and stabilize emulsion at a low pH [101]. Skjak-Bræk et al. [105] suggested inhomogeneity of low-molecular-weight alginate in gel formulation with low concentration of Ca^{+2} and absence of non-gelling ions Na^{+} .

Table 1. Skin-benefiting activities of polysaccharides derived from marine macroalgae.

No.	Species	Cosmetics Properties and/or Products	References
1.	<i>Ecklonia cava</i>	Anticoagulant activity	[106]
2.	<i>Ishige okamurae</i> , <i>Schizymenia dubyi</i> , <i>Ecklonia cava</i> , <i>Ecklonia stolonifera</i> , <i>Sargassum silauastrum</i>	Tyrosinase inhibition	[107–109]
3.	<i>Sargassum fusiforme</i> (formerly <i>Hizikia fusiformis</i>)	Collagenase and elastase inhibition	[110]
4.	<i>Saccharina japonica</i> (formerly <i>Laminaria japonica</i>)	Antioxidant activity	[111]
5.	<i>Neoporphyra haitanensis</i> (formerly <i>Porphyra haitanensis</i>), <i>Ulva australis</i> (formerly <i>Ulva pertusa</i>), <i>Ulva linza</i> (formerly <i>Enteromorpha linza</i>), <i>Bryopsis plumosa</i>	Antioxidant activity	[112]
6.	<i>Sargassum</i> sp., <i>Neopyropia yezoensis</i> (formerly <i>Porphyra yezoensis</i>)	Antilipidemic activity	[113,114]
7.	<i>Fucus</i> sp., <i>Laminaria</i> sp., <i>Sargassum</i> sp.	Skin-whitening effect	[115–117]
8.	<i>Turbinaria ornata</i>	Antioxidant, anti-inflammatory	[118]

Table 1. Cont.

No.	Species	Cosmetics Properties and/or Products	References
9.	<i>Sargassum polycystum</i>	Tyrosinase inhibition	[115]
10.	<i>Corallina pilulifera</i> , <i>Ecklonia cava</i>	Inhibition of MMP-2,9	[81,119]
11.	<i>Schizymenia binderi</i>	Antiviral activity	[120]
12.	<i>Fucus vesiculosus</i> , <i>Turbinaria conoides</i>	Antioxidant (photoprotection)	[121]
13.	<i>Gongolaria nodicaulis</i> (formerly <i>Cystoseira nodicaulis</i>), <i>Eisenia bicyclis</i> , <i>Ecklonia cava</i> subsp. <i>kurome</i> (formerly <i>Ecklonia kurome</i>)	Hyaluronidase inhibition Antiaging	[122,123]
14.	<i>Undaria pinnatifida</i> , <i>Codium tomentosum</i> , <i>Durvillaea antarctica</i> , <i>Saccharina japonica</i> (formery <i>Laminaria japonica</i>), <i>Ulva</i> sp.	Moisture retention	[120,124,125]
15.	<i>Sargassum patens</i>	Antiviral activity	[126]
16.	<i>Lobophora variegata</i>	Antiviral activity	[127]
17.	<i>Sargassum vulgare</i> , <i>Colpomenia sinuosa</i> , <i>Dictyopteris polypodioides</i> (formerly <i>Dictyopteris membranacea</i>)	Antimicrobial	[128]
18.	<i>Padina pavonica</i> , <i>Ecklonia cava</i>	Antimicrobial activity	[129]
19.	<i>Corallina</i> sp., <i>Saccharina japonica</i> (formerly <i>Laminaria japonica</i>)	Antimicrobial Moisturizer	[130,131]
20.	<i>Porphyra umbilicalis</i>	Reduced ROS by UV	[132,133]

There are a wide variety of polysaccharides that are useful in skin cosmetics, such as agar, alginic acid, carrageenan, porphyrin, laminarin, fucoidan, and ulvan. Many genera of agrophytes algae, such as *Gelidium* sp., *Gracilaria* sp., *Gelidiella* sp., *Pterocladia* sp., etc., are well-known producers of agar-agar [51,134]. Balboa et al. [135] suggested use of agar as a major ingredient in creams, as an emulsifier, stabilizer, moisturizer as well as in different cosmetic products such as lotion, deodorants, antiaging treatment, exfoliant, acne treatment, etc. Like agar, alginic acid is derived from several brown algal species (Fucales, Laminariales, *Ascophyllum* sp., *Durvillaea* sp., *Ecklonia* sp., *Laminaria* sp., *Macrocystis* sp., *Saccharina* sp., *Sargassum* sp., and *Turbinaria* sp.) [136–138]. Mafinowska [139] and Fabrowska et al. [140] reported its application in the formulation of skin-protective or barrier creams for the treatment of dermatitis, as well as suitable ingredients of beauty masks or facial packs. In addition, Kappa-, Iota-, Lambda-, Beta-carrageenan are extracted from several carrageenophytes, i.e., *Betaphycus gelatinum*, *Chondrus crispus*, *Euclima denticulatum*, *Gigartina* sp., *Kappaphycus alvarezii*, *Hypnea musciformis*, *Mastocarpus* sp., and *Mazzaella* sp., from the Rhodophyta. It is used in cosmetology for various applications, such as lotion, sun-ray protectors, medicines, deodorant sticks, sprays, and foams [141–144].

Moreover, porphyrin is a well-studied class of sulfated polysaccharides obtained from the aqueous extract of red algae *Porphyra* sp. and *Bangia* sp. [145,146]. It has shown potential cosmeceutical applications, such as skin-whitening, antiulcer, analgesic, and anti-inflammatory properties. Many brown seaweed species, including *Laminaria* sp., *Saccharina* sp., *Ascophyllum* sp., *Fucus* sp., *Sargassum* sp., and *Undaria* sp., are well-known for laminaran properties, such as antitumor, anti-inflammatory, antiviral, antioxidant, anticoagulant, and anti-cellulite properties [147–149]. Among all, sulfated polysaccharides have attractive considerable attention in cosmeceutical activities: UV protector, anti-inflammatory, antico-

agulant antithrombotic, tyrosinase inhibitor, antitumoral, antibacterial, antidiabetic, and antioxidative [150–153]. Moon et al. [154] found the role of fucoidan in an inhibition of matrix metalloproteinase induced by UVB radiation. Accordingly, Senni et al. [155] suggested its potential role in prevention of photoaging of the skin. Consequently, fucoidan acts as an inhibitor of tyrosinase and reduces skin pigmentation, while used in skin-whitening formulation [156,157]. Pereira [158], Carvalho and Pereira [159], and Geszteszi et al. [160] suggested ulvan as desirable raw material for cosmeceuticals. Apart from, ulvans have beneficial moisturizing, protective, antitumor, and antioxidative properties in gel formulation [161,162]. Yaich et al. [163] described the skin protective and bioactive effects of rhamnose and fucose against skin aging. The unique chemical and physiochemical properties of polysaccharides make them attractive candidates for novel functional and biologically active polymer for cosmeceuticals [134].

1.3. Proteins

Biological macromolecule protein is a polymer of amino acids that is present in all living organisms. It is a basic building block of almost all cellular processes. It may present itself in the form of enzymes, hormones, vitamins, and pigments [164,165]. Moreover, macroalgae contain different types of aliphatic amino acids, hydroxyl-group-containing amino acid, aromatic amino acid, mycosporine amino acids, etc., which are summarized in Table 2. In addition, different species of *Palmaria palmata*, *Chondrus crispus*, *Porphyra* sp. (Rhodophyta), *Undaria pinnatifida* (Phaeophyceae), *Ulva* sp. (Chlorophyta), and *Euchema* sp. (Rhodophyta) are reported for the quantity of amino acids they contain [166–168]. It is widely applicable in cosmeceutical preparation as a functional part. It exhibits many cosmeceutical activities, such as anti-inflammatory, antiaging, antioxidant, and photoprotection activities [169,170]. According to Fabrowska et al. [171], protein showed a moisturizing effect on human skin. MAAs (mycosporine-like amino acids) play their role in the absorption of solar energy that beneficiary in photoaging, as well as photo-damaging protection. There are different roles of MAAs, such as in UV protection, anti-photoaging, antioxidant, and anti-hypertensive activities, reported by researchers [172–175]. Moreover, Dunalp and Yamamoto [176] reported the importance of mycosporine amino acids (MAAs) as sunscreens to reduce UV-induced damage. MAAs play a major role in protection against damage caused by sunlight. They acting as antioxidant molecules which scavenge toxic oxygen radicals and protect skin against UV-induced damage [177]. Furthermore, MAAs act as protective solutes of cells against salt stress, desiccation, and thermal stress [178].

Table 2. Skin-benefiting activities of proteins/amino acids derived from marine macroalgae.

No.	Species	Cosmetics Properties and/or Products	References
1.	<i>Laminaria digitata</i>	Lipolytic activity	[179]
2.	<i>Porphyra umbilicalis</i>	Anti-UVA Antioxidant	[180] [181]
3.	<i>Ecklonia cava</i>	Antioxidant, chelating agent, radical scavenger	[182]
4.	<i>Palmaria palmata</i> , <i>Neopyropia yezoensis</i> (formerly <i>Porphyra yezoensis</i>), <i>Ulva prolifera</i> (formerly <i>Enteromorpha prolifera</i>)	Antioxidant activity	[183]
5.	<i>Sargassum polycystum</i>	Anti-melanogenesis/skin-whitening effect	[184]
6.	<i>Pelvetia canaliculata</i>	Antioxidant, collagen synthesis,	[185]
7.	<i>Jania rubens</i>	Anti-skin-cancer	[186]
8.	<i>Porphyra umbilicalis</i>	Sunscreen	[187]
9.	<i>Scytosiphon lomentaria</i>	Antioxidant	[188]
10.	<i>Furcellaria lumbricalis</i> , <i>Fucus vesiculosus</i>	Anti-skin-aging	[189]
11.	<i>Acanthophora nayadiformis</i>	Antioxidant, radical scavengers	[190]

Table 2. Cont.

No.	Species	Cosmetics Properties and/or Products	References
12.	<i>Limnospira maxima</i> (formerly <i>Spirulina maxima</i>), <i>Ulva lactuca</i> , <i>Rhizoclonium riparium</i> var. <i>implexum</i> (formerly <i>Lola implexa</i>)	Anti-skin-aging	[191]
13.	<i>Palmaria palmate</i>	Moisturizer, natural sunscreen, antioxidant	[192]
14.	<i>Neopyropia elongata</i> (formerly <i>Porphyra rosengurtii</i>)	Photoprotective effects	[193]
15.	<i>Ecklonia stolonifera</i>	Inhibition of MMP	[194,195]

Galland-Irmouli et al. [196] and Samarakoon and Jeon [197] reported various skin-benefiting activities of protein and amino acids: anti-inflammatory, antioxidant, antitumor, antiaging, skin protective, and moisturizing effects in cosmetic products and the natural moisturizing factor in human skin. Houston [198] found *Ulva australis* to be a good source of essential amino acids, such as histidine and taurine. whereas Galland-Irmouli et al. [199], Pereira [200], and Martínez–Hernández et al. [201] have shown that red alga *Palmaria palmata* and *Himanthalia elongata* are a high source of serine, alanine, and glutamic acid. Reef et al. [202] and Pereira, [203] detected mycosporines, such as amino acids (MAAs), in different red macroalgae (Rhodophyta): *Chondrus crispus*, *Palmaria palmata*, *Gelidium* sp., *Porphyra* sp., *Gracillaria cornea*, *Asparagopsis armata*, *Grateloupia lanceola*, and *Curdiea* sp. Pereira, [203] showed role of MAAs as UV protectors and activators of cell proliferation in cosmetics.

The red alga *Porphyra rosengurtii*–derived mycosporine-like amino acids Porphyra-334 and Shinorine are isolated and found to be very photostable and photoprotective when exposed to radiation [204]. These MAAs both played a role in sunburn cell formation and to be protective after UV radiation and eliminate damaged cells [205]. This combination also used in treatment of prevention towards skinfold thickening in the epidermis/dermis of hypodermic of mice.

1.4. Phenolic Compounds

Marine macroalgae are richer in various phenolic compounds, such as catechins, flavonols, flavonolglycosides, phloroglucinol, gallic acid, epicatechin, pyrocatechol, galate, flavonoids, anthocyanins, stilbenes, lignans, and phenolic polymers [206,207]. These types of phenolic compounds revealed their effect on MMP (Matrix Metalloproteinase complex) inhibition, as well as the reduction of collagen degradation [208]. This research also reported algal-derived phenolic compounds helpful to suppress both the protein and gene expression of the MMP complex. Ryu et al. [209] suggested that *Corallina pilulifera* (Rhodophyta) can inhibit the expression of MMP-2 and MMP-9. Another phenolic compound, sargachromanol E, from *Sargassum horneri* (Phaeophyceae), expressed its effect on antiaging activity [210]. Porphyra 334, a mycosporine amino acid from *Phycocalidia vietnamensis* (formerly *Porphyra vietnamensis*) (Rhodophyta), showed UV-absorbing properties [211]. Catechin and some other phycocompounds, such as flavonoids, polyphenol, and carotenoids, showed ROS scavenging, downregulation of the mitogen-activated protein kinase (MAPK) pathway, inhibition of MMP, and the elevation of collagen production, giving them wider application in cosmetics [212,213]. Moreover, brown macroalgae *Ecklonia cava*-derived compounds, such as phlorotannins, exhibit skin whitening/antityrosinase effect, whereas zeaxanthin from the microalga *Nannochloropsis oculata* (Ochrophyta and Eustigmatophyceae) extracts showed skin-whitening activity [214]. The beneficial activities of seaweed-derived phenolic compounds for skin uses are illustrated in Table 3. Pavia and Brock [215] and Bravo [216] identified phenolic compounds such as phlorotannins and phloroglucinol (1,3,5-trihydroxybenzene) in different brown algal families, such as Alariaceae, Sargassaceae, and Fucaceae. Different algal species are evaluated for antioxidant activity by different methods, such as DPPH (2,2-diphenyl-1-picrylhydrazyl) radical scavenging activity, ferrous ion-chelating ability, and ORAC (Oxygen Radical Absorbance

Capacity) [217,218]. Ferreres et al. [219] and Sanjeewa et al. [220] reported anti-allergic, anti-wrinkle, and skin antiaging activities of phlorotannins, due to hyaluronidase inhibition activity. Jang et al. [221] studied tyrosinase inhibition and the whitening effect of phlorotannins from *Sargassum fusiforme* (*Hijikia fusiformis*). Phlorotannins, eckol, Fucols, Fucophorethols, Fuhalols, Phlorethols from *Corallina pilulifera* (red algae) have beneficial cosmetic properties: antiaging, antiphotodaging, antioxidant, anti-allergic, anti-inflammatory, tyrosinase inhibition, and hyaluronidase inhibition [222–225]. Moreover, phlorotannins are reported to be inhibitors of melanin synthesis, as well as being protective against UVB photodamage [226]. Likewise, Handelman [227] and Wang et al. [228] revealed the inhibitory effect *Ecklonia cava*-derived phlorotannins on melanin synthesis and protective effects on UV damage.

Table 3. Skin beneficial activities of phenolic compounds derived from marine macroalgae.

No.	Species	Potential Phenolic Compound/s Studied	Cosmetics Properties and/or Products	References
1	<i>Sargassum muticum</i> , <i>Ishige okamurae</i> , <i>Ecklonia cava</i> , <i>Polysiphonia morrowii</i> , <i>Dictyopteria undulata</i> , <i>Sargassum micracanthum</i> , <i>Sargassum macrocarpum</i>	Total phenolic compounds	Antioxidant	[229]
2	<i>Acetabularia ryukyuensis</i> , <i>Undaria pinnatifida</i> , <i>Gelidium elegans</i>	Flavonoid	Antioxidant	[230]
3	<i>Sargassum siliquastrum</i>	Total phenolic compounds	Antioxidant	[231]
4	<i>Ascophyllum nodosum</i>	Phlorotannin Flavonoid	Antioxidant	[232,233]
5	<i>Ulva prolifera</i> (formerly <i>Enteromorpha prolifera</i>)	Flavonoid	Anti-inflammatory, Antiviral, Anticoagulant	[234]
6	<i>Fucus serratus</i> , <i>Sargassum muticum</i> , <i>Saccharina latissima</i> , <i>Laminaria digitata</i> , <i>Agarophyton vermiculophyllum</i> (formerly <i>Gracilaria vermiculophylla</i>)	Phenolic compound	Antioxidant	[235]
7	<i>Anthophycus longifolius</i> , <i>Sargassum plagiophyllum</i> , <i>Sargassum myriocystum</i>	Total Phenolic compounds	Antioxidant	[236]
8	<i>Chaetomorpha antennina</i>	Phenolic compound	Antiviral, Antibacterial, Antifungal, Anticancer	[237]
9	<i>Fucus vesiculosus</i>	Polyphenol	Antioxidant	[238]
10	<i>Halimeda maculosa</i> , <i>Halimeda opuntia</i>	Phenolic acid (compound)	Antioxidant	[239]
11	<i>Gongolaria barbata</i> (formerly <i>Cystoseira barbata</i>), <i>Scytosiphon lomentaria</i> , <i>Chondracanthus acicularis</i> (formerly <i>Gigartina acicularis</i>)	Phenolic compounds Carotenoids Flavonoids	Antioxidant	[240]
12	<i>Halimeda monile</i>	Phenolic acid	Antioxidative	[241]
13	<i>Fucus vesiculosus</i> , <i>Ascophyllum nodosum</i> , <i>Fucus serratus</i>	Polyphenols Phlorotannins	Antioxidant	[242]

Table 3. Cont.

No.	Species	Potential Phenolic Compound/s Studied	Cosmetics Properties and/or Products	References
14	<i>Sirophysalis trinodis</i> (formerly <i>Sirophysalis trinodis</i>)	Phlorotannins	Antioxidant	[243]
15	<i>Ecklonia cava</i>	Phloroglucinol	Antioxidant	[244]
16	<i>Fucus vesiculosus</i> , <i>Ascophyllum nodosum</i> , <i>Fucus serratus</i> ,	Phlorotannins	Antioxidant	[245]
17	<i>Ceramium rubrum</i> , <i>Cladophora vagabunda</i> , <i>Ulva intestinalis</i> (formerly <i>Enteromorpha intestinalis</i>)	Phenolic compound	Antioxidant	[246]
18	<i>Ulva lactuca</i>	Phlorotannins	Antioxidant	[247]
19	<i>Sargassum fusiforme</i> (formerly <i>Hizikia fusiformis</i>), <i>Ishige foliacea</i>	Phlorotannins	Tyrosinase inhibition, Antioxidant, Anti-Inflammatory, Anti-allergic	[248,249]
20	<i>Ecklonia stolonifera</i> , <i>Eisenia bicyclis</i>	Phlorotannins	Anti-inflammatory, Antioxidative	[250]
21	<i>Vertebrata thuyoides</i> (formerly <i>Boergeseniella thuyoides</i>), <i>Gracilaria multipartita</i>	Phenolic compound and Flavonoids	Antioxidant	[251]
22	<i>Sargassum pacificum</i> (formerly <i>Sargassum mangarevense</i>), <i>Turbinaria ornata</i>	Phenolic compound	Antioxidant, Antimicrobial	[252]
23	<i>Cladophora rupestris</i> , <i>Codium fragile</i>	Phenolic compound	Antioxidant, Mineralogenic	[253]
24	<i>Sargassum siliquastrum</i>	Fucoxanthin	Antioxidant	[254]
25	<i>Desmarestia ligulata</i> , <i>Dictyota kunthii</i> , <i>Laurencia chilensis</i> , <i>Chondracanthus chamissoi</i>	Flavonoids Polyphenols	Antioxidant, Cytotoxic, Anticancer	[255]
26	<i>Sargassum pacificum</i> (formerly <i>Sargassum mangarevense</i>), <i>Turbinaria ornata</i>	Phenolic compound	Antioxidant, Antimicrobial	[254]
27	<i>Ericaria selaginoides</i> (<i>Cystoseira tamariscifolia</i>)	Phenolic compound	Photoprotection	[256]
28	<i>Eisenia arborea</i>	Phlorotannins	Anti-inflammatory	[257]
29	<i>Pyropia columbina</i> (formerly <i>Porphyra columbina</i>)	Phenolic compound	Antioxidant	[258]

1.5. Pigments

Macroalgae is cultivated in a controlled condition to regulate the production of bioactive compounds such as phenolic compounds, pigments, carbohydrates, proteins, amino acids, vitamins, and minerals [259]. These algae-based valuable bioactive constituents gained attention in cosmeceutical activities [260]. This algae-derived metabolite can repair early signs of skin-aging, has an anti-wrinkle effect, exerts tightening effects, collagen synthesis, etc., as reported from *Arthrospira* species (Cyanobacteria) and *Chlorella vulgaris* (Chlorophyta) [261,262]. Marine algae contain a broad range of photosynthetic pigments chlorophylls, carotenoids (carotenes, xanthophylls, fucoxanthin, and peridinin), and phycobilins (phycocyanin and phycoerythrin) [263,264]. As suggested by many researchers, red algae contain chlorophyll, phycobilin, carotenoids, β carotene, lutein, phycocyanin, and

phycoerythrin Whereas brown algae possess chlorophyll a, c, carotenoids, fucoxanthin, and other pigments. Likewise, Chlorophyta revealed the presence of chlorophyll-a, -b, and -c and carotenoids [265–271]. Due to the richness of diversified pigments' profile, it is applied in various applications, such as photoprotection, anti-inflammatory effects, anticancer effects, and the inhibition of cell proliferation [272–276]. The benefits of seaweed-derived pigments are summarized in Table 4. According to Takaichi S. [277], Quilodrán et al. [278], and Amon and French [279], algae species are considered as a major source of β -carotene; likewise, some compounds, such as carotenoids, astaxanthin, and docosahexaenoic acid (DHA), show antioxidant activity. Hosikian et al. [280] evaluated the role of green photo-synthetic pigments in cosmetic industrial applications for antioxidant and antimutagenic properties. Spears [281] and La-Mer [282] suggested role of chlorophyll as natural coloring agents, deodorizing and antibacterial properties. In addition, these chlorophylls have high antioxidant activity and the ability for tissue-growth stimulation, making them useful to the cosmetic industry [283,284].

Carotenoids are widely applicable as natural dyes and antioxidants with antitumor, anti-inflammatory, and radical sequestering benefits [285–287]. They modulate UVA-induced gene expression and protect the skin against UV light [288]. Moreover, astaxanthin has a variety of roles in the prevention of UV-mediated photo-oxidation, tumors, and inflammation [289]. Additionally, fucoxanthin has protective effects on skin, making it consequently beneficial in cosmetics [290,291] Likewise, Kushwaha et al. [292] and Morabito et al. [282] reported carotenoids having antioxidant and anti-inflammatory properties that help for photoprotection and against UVA-damaging effects.

Table 4. Skin beneficial activities of marine macroalgae's pigments.

No.	Species	Potential Pigment/s Studied	Cosmetics Properties and/or Products	References
1	<i>Chaetomorpha antennina</i> , <i>Padina gymnospora</i>	Chlorophyll, Carotenoid, Xanthophylls, Antioxidant	Photoprotection	[283]
2	<i>Sargassum aquifolium</i> (formerly <i>Sargassum binderi</i>),	Fucoidan	Photoprotection	[284,285]
3	<i>Ulva lactuca</i> , <i>Caulerpa racemosa</i> , <i>Bryopsis plumosa</i> , <i>Gelidiella acerosa</i> , <i>Hypnea valentiae</i>	Chlorophyll Carotenoid	Photoprotection	[286]
4	<i>Sargassum ilicifolium</i>	Fucoxanthin	Photoprotection Antioxidant	[287]
5	<i>Sargassum polycistum</i>	Fucoxanthin β carotene α carotene	Antioxidant	[288]
6	<i>Haematococcus lacustris</i> (formerly <i>Haematococcus pluviialis</i>)	Lutein β carotene	Photo-oxidative	[289]
7	<i>Sacharina latissima</i> (formerly <i>Laminaria</i> <i>saccharina</i>)	Chlorophyll	Photo-inhibition	[290]
8	<i>Chondrus crispus</i>	Carotenoid	Photoprotection	[291]
9	<i>Kappaphycus alvarezii</i> , <i>Padina australis</i>	Chlorophyll a β carotene Fucoxanthin Zeaxanthin	Photoprotection	[292]

2. Discussion

Cosmetic researchers have focused their attention on marine organisms as an additional source of novel and useful natural ingredients. Diversified marine-algae-derived secondary metabolites are structurally more complex, with unique functionalities and properties. This review surveyed the potential applications of marine-algae-derived compounds for various skin benefits in the cosmetic industry. Though many seaweeds are exploited for their cosmetic properties, the research work on them is still incomplete, and so many species, either in full or in part, have not been explored. Hence, the cost-effective and efficient alternative standardized method to extract the bioactive phyco-constituents with significant productivity and activity is in growing demand. In future perspectives, the responsible molecular mechanism and safety concerns of these compounds are very important for future challenges in cosmeceuticals. Therefore, further investigations to study the precise molecular basis for the beneficial activity of marine algal components should be undertaken. Recently, *in silico* tools and techniques have been used to select functional materials derived from natural resources quickly and to predict the mechanisms of actions. Thus, this approach will be a helpful strategy for finding and understanding more effective compounds with the novel property.

3. Conclusions

The overexposure of human skin to different environmental stresses, such as pollutants and sun radiation, as well as chemical cosmeceutical ingredients—it increases the production of reactive oxygen species (ROS)—leads to many skin-damaging problems, such as aging, dullness, carcinogenesis, wrinkles, age spots, dark circles, etc. Marine-algae-based bioactive purified compounds demonstrated highly significant beneficiary applications in cosmetic formulas, as multiple functions, where they can be natural active constituents to the synthetic ingredients. Under different environmental factors, marine algae have the biosynthesis of primary and secondary metabolites for their survival. These biologically active constituents can be used as an active ingredient in the cosmetic industries due to their various skin benefits. It could be used as an antioxidant, antimicrobials, antibacterial, whitening agent, antiaging, anti-wrinkle, anti-acne, moisturizing, UV protection, deodorizing, anti-allergic, anti-inflammatory, sensory enhancer, viscosifying, stabilizer, and also for thickening in cosmetic industries. Sustainable use of marine algae and marine-algae-based molecules is crucial for humankind. Moreover, there are many cosmeceutical industries that already use extracts of marine algae and compounds in the formulation of many products. However, the monitoring of its biochemical profile presents a problem that needs to overcome. This can be solved by the development of seaweed cultivation and green extraction methods that are being analyzed with promising research results. However, many cosmetic companies' collaboration at the national and international level can improve the analytical methods of its screening for safety, thus enhancing consumer's safety towards marine-algae-based bioactive compounds in the cosmetic products. All mentioned marine algae in this review, possessing various bioactivities, are considered and utilized as a natural inexhaustible source for different cosmeceutical benefits.

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References

1. Millikan, L.E. Cosmetology, cosmetics, cosmeceuticals: Definitions and regulations. *Clin. Dermatol.* **2001**, *19*, 371–374. [[CrossRef](#)]
2. Jesumani, V.; Du, H.; Aslam, M.; Pei, P.; Huang, N. Potential Use of Seaweed Bioactive Compounds in Skincare—A Review. *Mar. Drugs* **2019**, *17*, 688. [[CrossRef](#)]
3. Agatonovic-Kustrin, S.; Morton, D. Cosmeceuticals derived from bioactive substances found in marine algae. *Oceanogr. Mar. Res.* **2013**, *1*, 2–11.
4. Guillerme, J.-B.; Couteau, C.; Coiffard, L. Applications for Marine Resources in Cosmetics. *Cosmetics* **2017**, *4*, 35. [[CrossRef](#)]
5. Łopaciuk, A.; Łoboda, M. Global beauty industry trends in the 21st century. In Proceedings of the Management, Knowledge and Learning International Conference, Celje, Slovenia, 20–22 June 2012; pp. 19–21.
6. Priyan Shanura Fernando, I.; Kim, K.N.; Kim, D.; Jeon, Y.J. Algal polysaccharides: Potential bioactive substances for cosmeceutical applications. *Crit. Rev. Biotechnol.* **2019**, *39*, 99–113. [[CrossRef](#)] [[PubMed](#)]
7. Gao, X.-H.; Zhang, L.; Wei, H.; Chen, H.-D. Efficacy and safety of innovative cosmeceuticals. *Clin. Dermatol.* **2008**, *26*, 367–374. [[CrossRef](#)]
8. Athukorala, Y.; Kim, K.-N.; Jeon, Y.-J. Antiproliferative and antioxidant properties of an enzymatic hydrolysate from brown alga, *Ecklonia cava*. *Food Chem. Toxicol.* **2006**, *44*, 1065–1074. [[CrossRef](#)] [[PubMed](#)]
9. Wijesinghe, W.A.J.P.; Jeon, Y.-J. Biological activities and potential cosmeceutical applications of bioactive components from brown seaweeds: A review. *Phytochem. Rev.* **2011**, *10*, 431–443. [[CrossRef](#)]
10. Nohynek, G.J.; Antignac, E.; Re, T.; Toutain, H. Safety assessment of personal care products/cosmetics and their ingredients. *Toxicol. Appl. Pharmacol.* **2010**, *243*, 239–259. [[CrossRef](#)] [[PubMed](#)]
11. Khan, A.D.; Alam, M.N. Cosmetics and their associated adverse effects: A review. *J. Appl. Pharm. Sci. Res.* **2019**, *2*, 1–6. [[CrossRef](#)]
12. Schrader, T.J.; Cooke, G.M. Examination of selected food additives and organochlorine food contaminants for androgenic activity in vitro. *Toxicol. Sci.* **2000**, *53*, 278–288. [[CrossRef](#)]
13. Zhang, Y.; De Sanjose, S.; Bracci, P.M.; Morton, L.M.; Wang, R.; Brennan, P.; Hartge, P.; Boffetta, P.; Becker, N.; Maynadie, M.; et al. Personal Use of Hair Dye and the Risk of Certain Subtypes of Non-Hodgkin Lymphoma. *Am. J. Epidemiol.* **2008**, *167*, 1321–1331. [[CrossRef](#)] [[PubMed](#)]
14. Health Canada. Report on human biomonitoring of environmental chemicals in Canada. In *Results of the Canadian Health Measures Survey Cycle 1 (2007–2009)*; Health Canada: Ottawa, ON, Canada, 2010; ISBN 978-1-100-15618-7.
15. Turkoglu, M.; Pekmezci, E.; Sakr, A. Evaluation of irritation potential of surfactant mixtures. *Int. J. Cosmet. Sci.* **1999**, *21*, 371–382. [[CrossRef](#)]
16. Bridges, B. Fragrance: Emerging health and environmental concerns. *Flavour Fragr. J.* **2002**, *17*, 361–371. [[CrossRef](#)]
17. Ulrich, G.; Schmutz, J.L.; Trechot, P.; Commun, N.; Barbaud, A. Sensitization to petrolatum: An unusual cause of false-positive drug patch-tests. *Allergy* **2004**, *59*, 1006–1009. [[CrossRef](#)] [[PubMed](#)]
18. Ratner, B.D.; Hoffman, A.S.; Schoen, F.J.; Lemons, J.E. *Biomaterials Science: An Introduction to Materials in Medicine*; Elsevier: Amsterdam, The Netherlands, 2004.
19. Ramakant, S.; Poornima, S.; Sapina, J.; Mathur, H.B.; Agarwal, H.C. Heavy metal in cosmetics. *Cent. Sci. Environ.* **2014**, *45*, 3–28.
20. Takizawa, T.; Imai, T.; Onose, J.-I.; Ueda, M.; Tamura, T.; Mitsumori, K.; Izumi, K.; Hirose, M. Enhancement of Hepatocarcinogenesis by Kojic Acid in Rat Two-Stage Models after Initiation with *N*-bis(2-hydroxypropyl)nitrosamine or *N*-diethylnitrosamine. *Toxicol. Sci.* **2004**, *81*, 43–49. [[CrossRef](#)] [[PubMed](#)]
21. Zeleke, A.T.; Alemu, Z.A. Determinants of under-five childhood diarrhea in Kotebe Health Center, Yeka Sub City, Addis Ababa, Ethiopia: A case-control study. *Glob. J. Med Res.* **2014**, *14*.
22. Dooms-Goossens, A. Cosmetics as causes of allergic contact dermatitis. *Cutis* **1993**, *52*, 316.
23. Kothalawala, S.G.; Chathurangi, D.U.; Yatiwella, L.N.S.B. Brief Overview of Bioactive Compounds in Seaweeds, Their Properties and Practical Applications in Functional Foods. *Int. J. Sci. Res. Publ. (IJSRP)* **2018**, *8*, 594–598. [[CrossRef](#)]
24. Siahaan, E.A.; Pangestuti, R.; Munandar, H.; Kim, S.-K. Cosmeceuticals Properties of Sea Cucumbers: Prospects and Trends. *Cosmetics* **2017**, *4*, 26. [[CrossRef](#)]
25. Ahmed, A.B.; Adel, M.; Karimi, P.; Peidayesh, M. Pharmaceutical, cosmeceutical, and traditional applications of marine carbohydrates. *Adv. Food Nutr. Res.* **2014**, *73*, 197–220. [[PubMed](#)]
26. Heo, S.-J.; Ko, S.-C.; Cha, S.-H.; Kang, D.-H.; Park, H.-S.; Choi, Y.-U.; Kim, D.; Jung, W.-K.; Jeon, Y.-J. Effect of phlorotannins isolated from *Ecklonia cava* on melanogenesis and their protective effect against photo-oxidative stress induced by UV-B radiation. *Toxicol. Vitro.* **2009**, *23*, 1123–1130. [[CrossRef](#)] [[PubMed](#)]
27. Pereira, L. Seaweeds as source of bioactive substances and skincare therapy—Cosmeceuticals, algotherapy, and thalasso-therapy. *Cosmetics* **2018**, *5*, 68. [[CrossRef](#)]
28. Kim, S.-K.; Ravichandran, Y.D.; Khan, S.B.; Kim, Y.T. Prospective of the cosmeceuticals derived from marine organisms. *Biotechnol. Bioprocess Eng.* **2008**, *13*, 511–523. [[CrossRef](#)]
29. Uppangala, N. Seaweeds show anti-cancer activity: Alternative cancer therapy. *Publish Biotechnology Articles or Industry News*, 12 June 2010.

30. Brown, E.; Allsopp, P.J.; Magee, P.; Gill, C.I.R.; Nitecki, S.; Strain, C.R.; McSorley, E.M. Seaweed and human health. *Nutr. Rev.* **2014**, *72*, 205–216. [[CrossRef](#)]
31. Shannon, E.; Abu-Ghannam, N. Seaweeds as nutraceuticals for health and nutrition. *Phycologia* **2019**, *58*, 563–577. [[CrossRef](#)]
32. Brownlee, I.A.; Allen, A.; Pearson, J.P.; Dettmar, P.W.; Havler, M.E.; Atherton, M.R.; Onsøyen, E. Alginate as a source of dietary fiber. *Crit. Rev. Food Sci. Nutr.* **2005**, *45*, 497–510. [[CrossRef](#)]
33. Pereira, L. *Guia Ilustrado das Macroalgas*; Coimbra University Press: Coimbra, Portugal, 2009.
34. Pereira, L. Cytological and cytochemical aspects in selected carrageenophytes (Gigartinales, Rhodophyta). *Adv. Algal Cell Biol.* **2012**, *19*, 81–104.
35. González-Minero, F.J.; Bravo-Díaz, L. The use of plants in skin-care products, cosmetics and fragrances: Past and present. *Cosmetics* **2018**, *5*, 50. [[CrossRef](#)]
36. Kalasariya, H.S.; Dave, M.P.; Yadav, V.K.; Patel, N.B. Beneficial effects of marine algae in skin moisturization and photoprotection. *Int. J. Pharm. Sci. Health CARE* **2020**, *5*, 1–11. [[CrossRef](#)]
37. Pal, A.; Kamthania, M.C.; Kumar, A. Bioactive Compounds and Properties of Seaweeds—A Review. *Open Access Libr. J.* **2014**, *1*, 1–17. [[CrossRef](#)]
38. Patel, N.B.; Tailor, V.; Rabadi, M.; Jain, A.; Kalasariya, H. Role of marine macroalgae in Skin hydration and photoprotection benefits: A review. *Int. J. Bot. Stud.* **2020**, *5*, 201–206.
39. Jain, A.; Patel, N.B.; Tailor, V.; Sathvara, S.; Kalasariya, H.S. An Appraisal on Antimicrobial applicability of Marine Macroalgae. *Int. Res. J. Eng. Technol.* **2020**, *7*, 735–739.
40. Nurjanah, N.M.; Anwar, E.; Luthfiyana, N.; Hidayat, T. Identification of bioactive compounds of seaweed *Sargassum* sp. and *Euचेuma cottonii* Doty as a raw sunscreen cream. *Proc. Pak. Acad. Sci. B Life Environ. Sci.* **2017**, *54*, 311–318.
41. Sahayaraj, K. Biological values and conservation of marine algae: An overview. In Proceedings of the Conservation and Sustainable Utilization of Marine Resources, National Conference on Conservation and Sustainable Utilization of Marine Resources, Tamil Nadu, India, 22–23 January 2015.
42. Kandale, A.; Meena, A.K.; Rao, M.M.; Panda, P.; Mangal, A.K.; Reddy, G.; Babu, R. Marine algae: An introduction, food value, and medicinal uses. *J. Pharm. Res.* **2011**, *4*, 219–221.
43. Anantharaman, P. Manual on the identification of seaweed. All India coordinate project on survey and inventorization of coastal and marine biodiversity. *J. Mar. Biol. Assoc. India.* **2002**, *29*, 1–9.
44. Aryee, A.N.; Agyei, D.; Akanbi, T.O. Recovery and utilization of seaweed pigments in food processing. *Curr. Opin. Food Sci.* **2018**, *19*, 113–119. [[CrossRef](#)]
45. Chojnacka, K.; Saeid, A.; Witkowska, Z.; Tuhy, L. Biologically active compounds in seaweed extracts—The prospects for the application. *Open Conf. Proc. J.* **2012**, *3*, 21–26. [[CrossRef](#)]
46. Gupta, S.; Abu-Ghannam, N. Bioactive potential and possible health effects of edible brown seaweeds. *Trends Food Sci. Technol.* **2011**, *22*, 315–326. [[CrossRef](#)]
47. Pereira, L. *Therapeutic and Nutritional Uses of Algae*; CRC Press: Boca Raton, FL, USA, 2018. [[CrossRef](#)]
48. Cotas, J.; Leandro, A.; Monteiro, P.; Pacheco, D.; Figueirinha, A.; Gonçalves, A.M.M.; Da Silva, G.J.; Pereira, L. Seaweed Phenolics: From Extraction to Applications. *Mar. Drugs* **2020**, *18*, 384. [[CrossRef](#)]
49. Indergaard, M. The aquatic resource. In *Biomass Utilization*; Springer: Boston, MA, USA, 1983; pp. 137–168.
50. Dias, V.; Bandeira, S.; Chauque, E.; Lipassula, M.; Mussagy, A. Evaluation of Phytocompounds and Chemical Elements Present in Selected Species of Seaweeds, to Sustain Future Quantitative Analysis for Bioactive Compounds. *J. Drug Deliv. Ther.* **2020**, *10*, 232–239. [[CrossRef](#)]
51. Malinowska, P. Algae extracts as active cosmetic ingredients. *Zesz. Nauk.* **2011**, *212*, 123–129.
52. Pereira, L.; Gheda, S.F.; Ribeiro-Claro, P.J.A. Analysis by Vibrational Spectroscopy of Seaweed Polysaccharides with Potential Use in Food, Pharmaceutical, and Cosmetic Industries. *Int. J. Carbohydr. Chem.* **2013**, *2013*, 1–7. [[CrossRef](#)]
53. Costa, L.; Fidelis, G.P.; Cordeiro, S.; Oliveira, R.; Sabry, D.; Câmara, R.; Nobre, L.; Costa, M.; Almeida-Lima, J.; Farias, E.; et al. Biological activities of sulfated polysaccharides from tropical seaweeds. *Biomed. Pharmacother.* **2010**, *64*, 21–28. [[CrossRef](#)]
54. Pereira, R.C.; Costa-Lotufo, L.V. Bioprospecting for bioactives from seaweeds: Potential, obstacles and alternatives. *Rev. Bras. Farm.* **2012**, *22*, 894–905. [[CrossRef](#)]
55. Bedoux, G.; Hardouin, K.; Burlot, A.S.; Bourgougnon, N. Bioactive components from seaweeds: Cosmetic applications and future development. *Adv. Bot. Res.* **2014**, *71*, 345–378.
56. Farage, M.A.; Miller, K.W.; Elsner, P.; Maibach, H.I. Intrinsic and extrinsic factors in skin ageing: A review. *Int. J. Cosmet. Sci.* **2008**, *30*, 87–95. [[CrossRef](#)]
57. Kang, S.-I.; Ko, H.-C.; Shin, H.-S.; Kim, H.-M.; Hong, Y.-S.; Lee, N.-H.; Kim, S.-J. Fucoxanthin exerts differing effects on 3T3-L1 cells according to differentiation stage and inhibits glucose uptake in mature adipocytes. *Biochem. Biophys. Res. Commun.* **2011**, *409*, 769–774. [[CrossRef](#)]
58. Lotti, T.; Ghersetich, I.; Grappone, C.; Dini, G. Proteoglycans in So-Called Cellulite. *Int. J. Dermatol.* **1990**, *29*, 272–274. [[CrossRef](#)]
59. Al-Bader, T.; Byrne, A.; Gillbro, J.; Mitarotonda, A.; Metois, A.; Vial, F.; Rawlings, A.V.; Laloëuf, A. Effect of cosmetic ingredients as anticellulite agents: Synergistic action of actives with in vitro and in vivo efficacy. *J. Cosmet. Dermatol.* **2012**, *11*, 17–26. [[CrossRef](#)]
60. Thomas, N.V.; Kim, S.-K. Beneficial Effects of Marine Algal Compounds in Cosmeceuticals. *Mar. Drugs* **2013**, *11*, 146–164. [[CrossRef](#)] [[PubMed](#)]

61. Nakamura, T.; Nagayama, K.; Uchida, K.; Tanaka, R. Antioxidant Activity of Phlorotannins Isolated from the Brown Alga *Eisenia bicyclis*. *Fish. Sci.* **1996**, *62*, 923–926. [[CrossRef](#)]
62. Chandrasekhar, S.; Esterman, M.A.; Hoffman, H.A. Microdetermination of proteoglycans and glycosaminoglycans in the presence of guanidine hydrochloride. *Anal. Biochem.* **1987**, *161*, 103–108. [[CrossRef](#)]
63. Sun, Y.; Chavan, M. Cosmetic Compositions Comprising Marine Plants. U.S. Patent 9,603,790, 28 March 2017.
64. Hagino, H.; Saito, M. Use of Algal Proteins in Cosmetics. European Patent EP1433463B1, 22 September 2010.
65. Leyton, A.; Pezoa-Conte, R.; Barriga, A.; Buschmann, A.; Mäki-Arvela, P.; Mikkola, J.-P.; Lienqueo, M. Identification and efficient extraction method of phlorotannins from the brown seaweed *Macrocystis pyrifera* using an orthogonal experimental design. *Algal Res.* **2016**, *16*, 201–208. [[CrossRef](#)]
66. Yu, P.; Gu, H. Bioactive Substances from Marine Fishes, Shrimps, and Algae and Their Functions: Present and Future. *Crit. Rev. Food Sci. Nutr.* **2015**, *55*, 1114–1136. [[CrossRef](#)]
67. De Jesus Raposo, M.F.; de Morais, A.M.B.; de Morais, R.M.C. Marine polysaccharides from algae with potential biomedical applications. *Mar Drugs* **2015**, *13*, 2967–3028. [[CrossRef](#)] [[PubMed](#)]
68. Kim, C.R.; Kim, Y.M.; Lee, M.K.; Kim, I.H.; Choi, Y.H.; Nam, T.J. Pyropiayezoensis peptide promotes collagen synthesis by activating the TGF- β /Smad signaling pathway in the human dermal fibroblast cell line Hs27. *Int. J. Mol. Med.* **2017**, *39*, 31–38. [[CrossRef](#)]
69. Speranza, L.; Pesce, M.; Patruno, A.; Franceschelli, S.; De Lutiis, M.A.; Grilli, A.; Felaco, M. Astaxanthin Treatment Reduced Oxidative Induced Pro-Inflammatory Cytokines Secretion in U937: SHP-1 as a Novel Biological Target. *Mar. Drugs* **2012**, *10*, 890–899. [[CrossRef](#)]
70. Dutta, P.K.; Dutta, J.; Tripathi, V.S. Chitin and chitosan: Chemistry, properties, and applications. *J. Sci. Ind. Res.* **2004**, *63*, 20–31.
71. Mandal, P.; Mateu, C.G.; Chattopadhyay, K.; Pujol, C.A.; Damonte, E.B.; Ray, B. Structural features and antiviral activity of sulfated fucans from the brown seaweed *Cystoseira indica*. *Antivir. Chem. Chemother.* **2007**, *18*, 153–162. [[CrossRef](#)]
72. Usov, A.I.; Zelinsky, N.D. Chemical structures of algal polysaccharides. In *Functional Ingredients from Algae for Foods and Nutraceuticals*; Woodhead Publishing: Sawston, UK, 2013; pp. 23–86.
73. Ahmed, A.; Taha, R. Marine Phytochemical Compounds and Their Cosmeceutical Applications. In *Marine Cosmeceuticals: Trends and Prospects*; Kim, S., Ed.; CRC Press: Boca Raton, FL, USA, 2011; pp. 51–61. [[CrossRef](#)]
74. Godlewska, K.; Michalak, I.; Tuhy, Ł.; Chojnacka, K. Plant Growth Biostimulants Based on Different Methods of Seaweed Extraction with Water. *BioMed Res. Int.* **2016**, *2016*, 5973760. [[CrossRef](#)]
75. Paduch, R.; Kandefer-Szerszeń, M.; Trytek, M.; Fiedurek, J. Terpenes: Substances useful in human healthcare. *Arch. Immunol. Ther. Exp.* **2007**, *55*, 315. [[CrossRef](#)] [[PubMed](#)]
76. Cha, S.H.; Ko, S.C.; Kim, D.; Jeon, Y.J. Screening of marine algae for potential tyrosinase inhibitor: Those inhibitors reduced tyrosinase activity and melanin synthesis in zebrafish. *J. Dermatol.* **2011**, *38*, 354–363. [[CrossRef](#)]
77. Murugan, K.; Iyer, V.V. Differential growth inhibition of cancer cell lines and antioxidant activity of extracts of red, brown, and green marine algae. *Vitr. Cell. Dev. Biol.-Anim.* **2013**, *49*, 324–334. [[CrossRef](#)] [[PubMed](#)]
78. Leandro, A.; Pereira, L.; Gonçalves, A.M.M. Diverse Applications of Marine Macroalgae. *Mar. Drugs* **2019**, *18*, 17. [[CrossRef](#)] [[PubMed](#)]
79. Buono, S.; Langelotti, A.L.; Martello, A.; Bimonte, M.; Tito, A.; Carola, A.; Apone, F.; Colucci, G.; Fogliano, V. Biological activities of dermatological interest by the water extract of the microalga *Botryococcus braunii*. *Arch. Dermatol. Res.* **2012**, *304*, 755–764. [[CrossRef](#)] [[PubMed](#)]
80. Kang, H.; Lee, C.H.; Kim, J.R.; Kwon, J.Y.; Seo, S.G.; Han, J.G.; Kim, B.G.; Kim, J.E.; Lee, K.W. *Chlorella vulgaris* attenuates *Dermatophagoides farinae*-induced atopic dermatitis-like symptoms in NC/Nga mice. *Int. J. Mol. Sci.* **2015**, *16*, 21021–21034. [[CrossRef](#)]
81. Pimentel, F.B.; Alves, R.C.; Rodrigues, F.; Oliveira, M.B.P.P. Macroalgae-Derived Ingredients for Cosmetic Industry—An Update. *Cosmetics* **2017**, *5*, 2. [[CrossRef](#)]
82. Kim, S.; You, D.H.; Han, T.; Choi, E.-M. Modulation of viability and apoptosis of UVB-exposed human keratinocyte HaCaT cells by aqueous methanol extract of laver (*Porphyra yezoensis*). *J. Photochem. Photobiol. B Biol.* **2014**, *141*, 301–307. [[CrossRef](#)] [[PubMed](#)]
83. Luthfiyana, N.; Hidayat, T.; Nurilmala, M.; Anwar, E. Utilization of seaweed porridge *Sargassum* sp. and *Euclima cottonii* as cosmetic in protecting the skin. *IOP Conf. Ser. Earth Environ. Sci.* **2019**, *278*, 012055.
84. Quah, C.C.; Kim, K.H.; Lau, M.S.; Kim, W.R.; Cheah, S.H.; Gundamaraju, R. Pigmentation and Dermal Conservative Effects of the Astonishing Algae *Sargassum polycystum* and *Padina tenuis* on Guinea Pigs, Human Epidermal Melanocytes (HEM) and Chang Cells. *Afr. J. Tradit. Complement. Altern. Med.* **2014**, *11*, 77–83. [[CrossRef](#)]
85. Maia Campos, P.M.; de Melo, M.O.; de Camargo, F.B., Jr. Effects of polysaccharide-based formulations on human skin. In *Polysaccharides*; Ramawat, K.G., Mérillon, J.-M., Eds.; Springer: Berlin/Heidelberg, Germany, 2014; pp. 1–8.
86. Kim, S.K. Marine cosmeceuticals. *J. Cosmet. Dermatol.* **2014**, *13*, 56–67. [[CrossRef](#)] [[PubMed](#)]
87. Jutur, P.P.; Nesamma, A.A.; Shaikh, K.M. Algae-Derived Marine Oligosaccharides and Their Biological Applications. *Front. Mar. Sci.* **2016**, *3*, 83. [[CrossRef](#)]
88. Venkatesan, J.; Anil, S.; Kim, S.K. Introduction to seaweed polysaccharides. In *Seaweed Polysaccharides*; Elsevier: Amsterdam, The Netherlands, 2017; pp. 1–9.

89. Percival, E. The polysaccharides of green, red and brown seaweeds: Their basic structure, biosynthesis and function. *Br. Phycol. J.* **1979**, *14*, 103–117. [[CrossRef](#)]
90. Song, Y.S.; Balcos, M.C.; Yun, H.Y.; Baek, K.J.; Kwon, N.S.; Kim, M.K.; Kim, D.S. ERK activation by fucoidan leads to inhibition of mel-anogenesis in Mel-Ab cells. *Korean J. Physiol. Pharm.* **2015**, *19*, 29–34. [[CrossRef](#)]
91. Shao, P.; Shao, J.; Han, L.; Lv, R.; Sun, P. Separation, preliminary characterization, and moisture-preserving activity of polysaccharides from *Ulva fasciata*. *Int. J. Biol. Macromol.* **2015**, *72*, 924–930. [[CrossRef](#)]
92. Fujimura, T.; Tsukahara, K.; Moriwaki, S.; Kitahara, T.; Takema, Y. Effects of natural product extracts on contraction and mechanical properties of fibroblast populated collagen gel. *Biol. Pharm. Bull.* **2000**, *23*, 291–297. [[CrossRef](#)]
93. Holtkamp, A.D.; Kelly, S.; Ulber, R.; Lang, S. Fucoidans and fucoidanases focus on techniques for molecular structure elucidation and modification of marine polysaccharides. *Appl. Microbiol. Biotechnol.* **2009**, *82*, 1–11. [[CrossRef](#)] [[PubMed](#)]
94. Fujimura, T.; Tsukahara, K.; Moriwaki, S.; Kitahara, T.; Sano, T.; Takema, Y. Treatment of human skin with an extract of *Fucus vesiculosus* changes its thickness and mechanical properties. *J. Cosmet. Sci.* **2002**, *53*, 1–9.
95. Teixeira, M.M.; Hellewell, P.G. The effect of the selectin binding polysaccharide fucoidin on eosinophil recruitment in vivo. *Br. J. Pharmacol.* **1997**, *120*, 1059–1066. [[CrossRef](#)]
96. Chen, Y.H.; Tu, C.J.; Wu, H.T. Growth-inhibitory effects of the red alga *Gelidium amansii* on cultured cells. *Biol. Pharm. Bull.* **2004**, *27*, 180–184. [[CrossRef](#)] [[PubMed](#)]
97. Campo, V.L.; Kawano, D.F.; da Silva, D.B., Jr.; Carvalho, I. Carrageenans: Biological properties, chemical modifications and structural analysis—A review. *Carbohydr. Polym.* **2009**, *77*, 167–180. [[CrossRef](#)]
98. Munaf, E.; Zein, R.; Dharm, A.; Lim, L.W.; Takeuchi, T. Optimization study of carrageenan extraction from red algae (*Euclima cottonii*). *J. Ris. Kim.* **2009**, *2*, 120–126. [[CrossRef](#)]
99. Podkorytova, A.V.; Vafina, L.H.; Kovaleva, E.A.; Mikhailov, V.I. Production of algal gels from the brown alga, *Laminaria japonica* Aresch., and their biotechnological applications. *J. Appl. Phycol.* **2007**, *19*, 827–830. [[CrossRef](#)]
100. Skjak-Bræk, G.; Grasdalen, H.; Smidsrød, O. Inhomogeneous polysaccharide ionic gels. *Carbohydr. Polym.* **1989**, *10*, 31–54. [[CrossRef](#)]
101. Jung, W.-K.; Athukorala, Y.; Lee, Y.-J.; Cha, S.H.; Lee, C.-H.; Vasanthan, T.; Choi, K.-S.; Yoo, S.-H.; Kim, S.-K.; Jeon, Y.-J. Sulfated polysaccharide purified from *Ecklonia cava* accelerates antithrombin III-mediated plasma proteinase inhibition. *Environ. Boil. Fishes* **2007**, *19*, 425–430. [[CrossRef](#)]
102. Paudel, P.; Wagle, A.; Seong, S.H.; Park, H.J.; Jung, H.A.; Choi, J.S. A New Tyrosinase Inhibitor from the Red Alga *Symphyclocladia latiuscula* (Harvey) Yamada (Rhodomelaceae). *Mar. Drugs* **2019**, *17*, 295. [[CrossRef](#)] [[PubMed](#)]
103. Manandhar, B.; Wagle, A.; Seong, S.H.; Paudel, P.; Kim, H.-R.; Jung, H.A.; Choi, J.S. Phlorotannins with Potential Anti-tyrosinase and Antioxidant Activity Isolated from the Marine Seaweed *Ecklonia stolonifera*. *Antioxidants* **2019**, *8*, 240. [[CrossRef](#)]
104. Kang, H.S.; Kim, H.R.; Byun, D.S.; Son, B.W.; Nam, T.J.; Choi, J.S. Tyrosinase inhibitors isolated from the edible brown alga *Ecklonia stolonifera*. *Arch. Pharm. Res.* **2004**, *27*, 1226–1232. [[CrossRef](#)]
105. Wang, L.; Lee, W.; Oh, J.Y.; Cui, Y.R.; Ryu, B.; Jeon, Y.J. Protective effect of sulfated polysaccharides from celluclast-assisted extract of *Hizikia fusiforme* against ultraviolet B-induced skin damage by regulating NF- κ B, AP-1, and MAPKs signaling pathways in vitro in human dermal fibroblasts. *Mar. Drugs* **2018**, *16*, 239. [[CrossRef](#)]
106. Rupérez, P.; Ahrazem, O.; Leal, J.A. Potential Antioxidant Capacity of Sulfated Polysaccharides from the Edible Marine Brown Seaweed *Fucus vesiculosus*. *J. Agric. Food Chem.* **2002**, *50*, 840–845. [[CrossRef](#)]
107. Zhang, Z.; Wang, F.; Wang, X.; Liu, X.; Hou, Y.; Zhang, Q. Extraction of the polysaccharides from five algae and their potential antioxidant activity in vitro. *Carbohydr. Polym.* **2010**, *82*, 118–121. [[CrossRef](#)]
108. Tsuge, K.; Okabe, M.; Yoshimura, T.; Sumi, T.; Tachibana, H.; Yamada, K. Dietary Effects of Porphyran from *Porphyra yezoensis* on Growth and Lipid Metabolism of Sprague-Dawley Rats. *Food Sci. Technol. Res.* **2004**, *10*, 147–151. [[CrossRef](#)]
109. Chen, S.; Wang, W.; Liu, H.; Li, C.; Liu, C. Purification and lowering hyperlipidemia activity of fucoidan from *Sargassum henslowianum*. *Shipin Yu Fajiao Gongye* **2010**, *36*, 28–31.
110. Fernando, I.S.; Sanjeewa, K.A.; Samarakoon, K.W.; Kim, H.S.; Gunasekara, U.K.; Park, Y.J.; Abeytunga, D.T.; Lee, W.W.; Jeon, Y.J. The potential of fucoidans from *Chnoospora minima* and *Sargassum polycystum* in cosmetics: Antioxidant, anti-inflammatory, skin-whitening, and antiwrinkle activities. *J. Appl. Phycol.* **2018**, *30*, 3223–3232. [[CrossRef](#)]
111. Wang, Z.-J.; Xu, W.; Liang, J.-W.; Wang, C.-S.; Kang, Y. Effect of fucoidan on B16 murine melanoma cell melanin formation and apoptosis. *Afr. J. Tradit. Complement. Altern. Med.* **2017**, *14*, 149–155. [[CrossRef](#)] [[PubMed](#)]
112. Yu, P.; Sun, H. Purification of a fucoidan from kelp polysaccharide and its inhibitory kinetics for tyrosinase. *Carbohydr. Polym.* **2014**, *99*, 278–283. [[CrossRef](#)]
113. Ananthi, S.; Raghavendran, H.R.; Sunil, A.G.; Gayathri, V.; Ramakrishnan, G.; Vasanthi, H.R. In vitro antioxidant and in vivo anti-inflammatory potential of crude polysaccharide from *Turbinaria ornata* (Marine Brown Alga). *Food Chem. Toxicol.* **2010**, *48*, 187–192. [[CrossRef](#)] [[PubMed](#)]
114. Kim, M.-M.; Van Ta, Q.; Mendis, E.; Rajapakse, N.; Jung, W.-K.; Byun, H.-G.; Jeon, Y.-J.; Kim, S.-K. Phlorotannins in *Ecklonia cava* extract inhibit matrix metalloproteinase activity. *Life Sci.* **2006**, *79*, 1436–1443. [[CrossRef](#)]
115. Joe, M.-J.; Kim, S.-N.; Choi, H.-Y.; Shin, W.-S.; Park, G.-M.; Kang, D.-W.; Kim, Y.K. The Inhibitory Effects of Eckol and Dieckol from *Ecklonia stolonifera* on the Expression of Matrix Metalloproteinase-1 in Human Dermal Fibroblasts. *Biol. Pharm. Bull.* **2006**, *29*, 1735–1739. [[CrossRef](#)] [[PubMed](#)]

116. Arora, N.; Agarwal, S.; Murthy, R.S. Latest technology advances in cosmeceuticals. *Int. J. Pharm. Sci. Drug Res.* **2012**, *4*, 168–182.
117. Wang, H.-M.D.; Li, X.-C.; Lee, D.-J.; Chang, J.-S. Potential biomedical applications of marine algae. *Bioresour. Technol.* **2017**, *244*, 1407–1415. [[CrossRef](#)] [[PubMed](#)]
118. Bu, H.J.; Ham, Y.M.; Kim, J.M.; Lee, S.J.; Hyun, J.W.; Lee, N.H. Elastase and hyaluronidase inhibition activities of phlorotannins isolated from *Ecklonia cava*. *Korean J. Pharm.* **2006**, *37*, 92–96.
119. Shibata, T.; Fujimoto, K.; Nagayama, K.; Yamaguchi, K.; Nakamura, T. Inhibitory activity of brown algal phlorotannins against hyaluronidase. *Int. J. Food Sci. Technol.* **2002**, *37*, 703–709. [[CrossRef](#)]
120. Choi, J.-S.; Moon, W.S.; Na Choi, J.; Hun, K.; Moon, S.H.; Cho, K.K.; Han, C.-J.; Choi, I.S. Effects of seaweed *Laminaria japonica* extracts on skin moisturizing activity in vivo. *J. Cosmet. Sci.* **2013**, *64*, 193–205.
121. Zhu, W.; Chiu, L.C.; Ooi, V.E.; Chan, P.K.; Ang, P.O., Jr. Antiviral property and mechanisms of a sulphated polysaccharide from the brown alga *Sargassum patens* against Herpes simplex virus type 1. *Phytomedicine* **2006**, *13*, 695–701. [[CrossRef](#)]
122. Queiroz, K.; Medeiros, V.; Queiroz, L.; Abreu, L.; Rocha, H.A.; Ferreira, C.; Jucá, M.; Aoyama, H.; Leite, E. Inhibition of reverse transcriptase activity of HIV by polysaccharides of brown algae. *Biomed. Pharmacother.* **2008**, *62*, 303–307. [[CrossRef](#)]
123. Khalil, A.M.; Daghaman, I.M. Fady, A.A. Antifungal Potential in Crude Extracts of Five Selected Brown Seaweeds Collected from the Western Libya Coast. *J. Microbiol. Mod. Tech.* **2015**, *1*, 103.
124. Lee, M.H.; Oh, S.M.; Chee, H.Y. Antifungal Activities of Dieckol Isolated from the Marine Brown Alga *Ecklonia cava* against *Trichophyton rubrum*. *J. Korean Soc. Appl. Biol. Chem.* **2010**, *53*, 504–507. [[CrossRef](#)]
125. Sebaaly, C.; Kassem, S.; Grishina, E.; Kanaan, H.; Sweidan, A.; Chmit, M.S.; Kanaan, H.M. Anticoagulant and antibacterial activities of polysaccharides of red algae *Corallina* collected from Lebanese coast. *J. Appl. Pharm. Sci.* **2014**, *4*, 30.
126. Wang, J.; Jin, W.; Hou, Y.; Niu, X.; Zhang, H.; Zhang, Q. Chemical composition and moisture-absorption/retention ability of poly-saccharides extracted from five algae. *Int. J. Biol. Macromol.* **2013**, *57*, 26–29. [[CrossRef](#)] [[PubMed](#)]
127. Ryu, J.; Park, S.-J.; Kim, I.-H.; Choi, Y.H.; Nam, T.-J. Protective effect of porphyra-334 on UVA-induced photoaging in human skin fibroblasts. *Int. J. Mol. Med.* **2014**, *34*, 796–803. [[CrossRef](#)] [[PubMed](#)]
128. Daniel, S.; Cornelia, S.; Fred, Z. UV-A sunscreen from red algae for protection against premature skin aging. *Cosmet Toilet. Manuf. Worldw.* **2004**, *2004*, 139–143.
129. Arad, S.; Levy-Ontman, O. Red microalgal cell-wall polysaccharides: Biotechnological aspects. *Curr. Opin. Biotechnol.* **2010**, *21*, 358–364. [[CrossRef](#)]
130. De Philippis, R.; Sili, C.; Paperi, R.; Vincenzini, M. Exopolysaccharide-producing cyanobacteria and their possible exploitation: A review. *Environ. Boil. Fishes* **2001**, *13*, 293–299. [[CrossRef](#)]
131. Balboa, E.M.; Conde, E.; Soto, M.L.; Pérez-Armada, L.; Domínguez, H. Cosmetics from marine sources. In *Springer Handbook of Marine Biotechnology*; Kim, S.-K., Ed.; Springer: Berlin/Heidelberg, Germany, 2015; pp. 1015–1042. ISBN 978-3-642-53970-1.
132. Pereira, L. Biological and therapeutic properties of the seaweed polysaccharides. *Int. Biol. Rev.* **2018**, *2*, 1–50. [[CrossRef](#)]
133. Sreekumar, K.; Bindhu, B. Alginic acid: A potential biopolymer from brown algae. *Mater. Int.* **2020**, *2*, 433–438.
134. Fabrowska, J.; Łeska, B.; Schroeder, G.; Messyas, B.; Pikosz, M. Biomass and extracts of algae as material for cosmetics. In *Marine Algae Extracts*; Kim, S.-K., Chojnacka, K., Eds.; Wiley-VCH, Verlag GmbH & Co. KGaA: Weinheim, Germany, 2015; pp. 681–706, ISBN 9783527337088.
135. Fernando, I.S.; Sanjeeva, K.A.; Kim, S.-Y.; Lee, J.-S.; Jeon, Y.-J. Reduction of heavy metal (Pb²⁺) biosorption in zebrafish model using alginic acid purified from *Ecklonia cava* and two of its synthetic derivatives. *Int. J. Biol. Macromol.* **2017**, *106*, 330–337. [[CrossRef](#)]
136. Pereira, L.; Amado, A.M.; Critchley, A.T.; van de Velde, F.; Ribeiro-Claro, P.J.A. Identification of selected seaweed polysaccharides (Phycocolloids) by vibrational spectroscopy (FTIR-ATR and FT-Raman). *Food Hydrocoll.* **2009**, *23*, 1903–1909. [[CrossRef](#)]
137. Charlier, R.H.; Chaineux, M.-C.P. The Healing Sea: A Sustainable Coastal Ocean Resource: Thalassotherapy. *J. Coast. Res.* **2009**, *254*, 838–856. [[CrossRef](#)]
138. Pereira, L. *ALGAE. Litoral of Viana do Castelo: Uses in Agriculture, Gastronomy and Food Industry (Bilingual)*; Câmara Municipal de Viana do Castelo: Viana do Castelo, Portugal, 2010; pp. 7–8, ISBN 978-972-588-218-4.
139. Gutiérrez, G. Compositions of Padina Algae or Their Extracts, and Their Pharmaceutical, Food Compositions, or Use for the Culture of Molluscs or Arthropods. European Patent EP 0655250 A1, 31 May 1995. Available online: <https://patents.google.com/patent/EP0655250A1/en> (accessed on 1 October 2018).
140. Villarroel, L.H.; Zanlungo, A.B. Structural studies on the porphyran from *Porphyra columbina* (Montagne). *Carbohydr. Res.* **1981**, *88*, 139–145. [[CrossRef](#)]
141. Lourenço-Lopes, C.; Fraga-Corral, M.; Jimenez-Lopez, C.; Pereira, A.G.; Garcia-Oliveira, P.; Carpena, M.; Prieto, M.A.; Simal-Gandara, J. Metabolites from Macroalgae and Its Applications in the Cosmetic Industry: A Circular Economy Approach. *Resources* **2020**, *9*, 101. [[CrossRef](#)]
142. Joshi, S.; Kumari, R.; Upasani, V.N. Applications of algae in cosmetics: An overview. *Int. J. Innov. Res. Sci. Eng. Technol.* **2018**, *7*, 1269–1278.
143. Kim, M.-S.; Oh, G.-H.; Kim, M.-J.; Hwang, J.-K. Fucosterol Inhibits Matrix Metalloproteinase Expression and Promotes Type-1 Procollagen Production in UVB-induced HaCaT Cells. *Photochem. Photobiol.* **2013**, *89*, 911–918. [[CrossRef](#)] [[PubMed](#)]
144. Lorbeer, A.J.; Tham, R.; Zhang, W. Potential products from the highly diverse and endemic macroalgae of Southern Australia and pathways for their sustainable production. *Environ. Boil. Fishes* **2013**, *25*, 717–732. [[CrossRef](#)]

145. Kuznetsova, T.A.; Besednova, N.N.; Mamaev, A.; Momot, A.P.; Shevchenko, N.M.; Zvyagintseva, T.N. Anticoagulant Activity of Fucoidan from Brown Algae *Fucus evanescens* of the Okhotsk Sea. *Bull. Exp. Biol. Med.* **2003**, *136*, 471–473. [[CrossRef](#)] [[PubMed](#)]
146. Wu, L.; Sun, J.; Su, X.; Yu, Q.; Yu, Q.; Zhang, P. A review about the development of fucoidan in antitumor activity: Progress and challenges. *Carbohydr. Polym.* **2016**, *154*, 96–111. [[CrossRef](#)] [[PubMed](#)]
147. Saravana, P.S.; Cho, Y.-N.; Patil, M.; Cho, Y.-J.; Kim, G.-D.; Park, Y.B.; Woo, H.-C.; Chun, B.-S. Hydrothermal degradation of seaweed polysaccharide: Characterization and biological activities. *Food Chem.* **2018**, *268*, 179–187. [[CrossRef](#)]
148. Moon, H.E.; Islam, N.; Ahr, B.R.; Chowdhury, S.S.; Sohn, H.S.; Jung, H.A.; Choi, J.S. Protein tyrosine phosphatase 1B and α -glucosidase inhibitory phlorotannins from edible brown algae *Ecklonia stolonifera* and *Eisenia bicyclis*. *Biosci. Biotechnol. Biochem.* **2011**, *75*, 1472–1480. [[CrossRef](#)]
149. Senni, K.; Gueniche, F.; Foucault-Bertaud, A.; Igondjo-Tchen, S.; Fioretti, F.; Collic-Jouault, S.; Durand, P.; Guezennec, J.; Godeau, G.; Letourneur, D. Fucoidan a sulfated polysaccharide from brown algae is a potent modulator of connective tissue proteolysis. *Arch. Biochem. Biophys.* **2006**, *445*, 56–64. [[CrossRef](#)] [[PubMed](#)]
150. Kim, K.B.; Jeong, S.M.; Kim, M.J.; Ahn, D.H. Tyrosinase Inhibitory Effects of Sargachromanol G, Sargachromanol I and Mojaban-chromanol b isolated from *Myagropsis myagroides*. *Indian J. Pharm. Sci.* **2020**, *82*, 170–173. [[CrossRef](#)]
151. Wijesinghea, W.A.J.P.; Jeona, Y.-J. Biological activities and potential industrial applications of fucose rich sulfated polysaccharides and fucoidans isolated from brown seaweeds: A review. *Carbohydr. Polym.* **2012**, *88*, 13–20. [[CrossRef](#)]
152. Morelli, A.; Massironi, A.; Puppi, D.; Creti, D.; Martinez, E.D.; Bonistalli, C.; Fabroni, C.; Morgenni, F.; Chiellini, F. Development of ulvan-based emulsions containing flavour and fragrances for food and cosmetic applications. *Flavour Fragr. J.* **2019**, *34*, 411–425. [[CrossRef](#)]
153. Carvalho, L.G.; Pereira, L. Review of marine algae as source of bioactive metabolites. In *Marine Algae—Biodiversity, Taxonomy, Environmental Assessment and Biotechnology*, 1st ed.; Pereira, L., Neto, J.M., Eds.; CRC Press: Boca Raton, FL, USA, 2015; ISBN 9781466581678.
154. Ukai, K.; Mizutani, Y.; Hisada, K.; Yokoyama, M.; Futaki, S.; Toya, H. Fuel Electrode Material, a Fuel Electrode, and a Solid Oxide Fuel Cell. U.S. Patent 20060110633 A1, 25 May 2006.
155. Cindana Mo'o, F.R.; Wilar, G.; Devkota, H.P.; Wathoni, N. Ulvan, a polysaccharide from macroalga *Ulva* sp.: A review of chemistry, biological activities and potential for food and biomedical applications. *Appl. Sci.* **2020**, *10*, 5488. [[CrossRef](#)]
156. Taher, P.M.; Ruslan, F.S.; Susanti, D.; Noor, N.M.; Aminudin, N.I. Bioactive Compounds, Cosmeceutical and Nutraceutical Applications of Green Seaweed Species (Chlorophyta). *Squalen Bull. Mar. Fish. Postharvest Biotechnol.* **2021**, *16*, 41–55. [[CrossRef](#)]
157. Yaich, H.; Ben Amira, A.; Abbes, F.; Bouaziz, M.; Besbes, S.; Richel, A.; Blecker, C.; Attia, H.; Garna, H. Effect of extraction procedures on structural, thermal and antioxidant properties of ulvan from *Ulva lactuca* collected in Monastir coast. *Int. J. Biol. Macromol.* **2017**, *105*, 1430–1439. [[CrossRef](#)]
158. Lahaye, M.; Robic, A. Structure and functional properties of ulvan, a polysaccharide from green seaweeds. *Biomacromolecules* **2007**, *8*, 1765–1774. [[CrossRef](#)]
159. Stengel, D.B.; Connan, S. Marine algae: A source of biomass for biotechnological applications. In *Natural Products from Marine Algae*; Humana Press: New York, NY, USA, 2015; pp. 1–37.
160. Černá, M. Seaweed proteins and amino acids as nutraceuticals. *Adv. Food Nutr. Res.* **2011**, *64*, 297–312.
161. Florence, J. Seaweed proteins: Biochemical, nutritional aspects and potential uses. *Trends Food Sci. Technol.* **1999**, *10*, 25–28. [[CrossRef](#)]
162. Harnedy, P.A.; Fitzgerald, R.J. Bioactive proteins and peptides from macroalgae, fish, shellfish, and marine processing waste. Marine proteins and peptides. *Mar. Proteins Pept. Biol. Act. Appl.* **2013**, *19*, 5–39.
163. Holdt, S.L.; Kraan, S. Bioactive compounds in seaweed: Functional food applications and legislation. *Environ. Biol. Fishes* **2011**, *23*, 543–597. [[CrossRef](#)]
164. Yuan, Y.V.; Westcott, N.D.; Hu, C.; Kitts, D.D. Mycosporine-like amino acid composition of the edible red alga, *Palmaria palmata* (dulse) harvested from the west and east coasts of Grand Manan Island, New Brunswick. *Food Chem.* **2009**, *112*, 321–328. [[CrossRef](#)]
165. Gerald, V.; Pinto, E. Mycosporine-Like Amino Acids (MAAs): Biology, Chemistry and Identification Features. *Pharmaceuticals* **2021**, *14*, 63. [[CrossRef](#)]
166. Lawrence, K.; Long, P.F.; Young, A.R. Mycosporine-Like Amino Acids for Skin Photoprotection. *Curr. Med. Chem.* **2019**, *25*, 5512–5527. [[CrossRef](#)] [[PubMed](#)]
167. Lee, J.H.; Kim, H.S.; Seo, H.H.; Song, M.Y.; Kulkarni, A.; Choi, Y.H.; Kim, K.W.; Moh, S.H. Antiaging effects of algae-derived mycosporine-like amino acids (MAAs) on skin. In *Textbook of Aging Skin*; Springer: Berlin/Heidelberg, Germany, 2015; Volume 5.
168. Dunlap, W.C.; Yamamoto, Y. Small-molecule antioxidants in marine organisms: Antioxidant activity of mycosporine-glycine. *Comp. Biochem. Physiol. Part B Biochem. Mol. Biol.* **1995**, *112*, 105–114. [[CrossRef](#)]
169. Pangestuti, R.; Shin, K.-H.; Kim, S.-K. Anti-Photoaging and Potential Skin Health Benefits of Seaweeds. *Mar. Drugs* **2021**, *19*, 172. [[CrossRef](#)]
170. Oren, A.; Gunde-Cimerman, N. Mycosporines and mycosporine-like amino acids: UV protectants or multipurpose secondary metabolites? *FEMS Microbiol.* **2007**, *269*, 1–10. [[CrossRef](#)]
171. Jang, W.S.; Choung, S.Y. Antiobesity Effects of the Ethanol Extract of *Laminaria japonica* Areshoung in High-Fat-Diet-Induced Obese Rat. *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 492807. [[CrossRef](#)] [[PubMed](#)]

172. Zhang, Q.; Li, N.; Liu, X.; Zhao, Z.; Li, Z.; Xu, Z. The structure of a sulfated galactan from *Porphyra haitanensis* and its in vivo anti-oxidant activity. *Carbohydr. Res.* **2004**, *339*, 105–111. [[CrossRef](#)]
173. Carreto, J.I.; Carignan, M.O. Mycosporine-Like Amino Acids: Relevant Secondary Metabolites. Chemical and Ecological Aspects. *Mar. Drugs* **2011**, *9*, 387–446. [[CrossRef](#)]
174. Fleurence, J.; Morançais, M.; Dumay, J. Seaweed proteins. In *Proteins in Food Processing*; Woodhead Publishing: Sawston, UK, 2018; pp. 245–262.
175. Farasat, M.; Khavari-Nejad, R.-A.; Nabavi, S.M.B.; Namjooyan, F. Antioxidant Activity, Total Phenolics and Flavonoid Contents of some Edible Green Seaweeds from Northern Coasts of the Persian Gulf. *Iran. J. Pharm. Res. IJPR* **2014**, *13*, 163–170.
176. Song, T.-Y.; Chen, C.-L.; Yang, N.-C.; Fu, C.-S.; Chang, Y.-T. The correlation of in vitro mushroom tyrosinase activity with cellular tyrosinase activity and melanin formation in melanoma cells A2058. *J. Food Drug Anal.* **2010**, *17*, 4. [[CrossRef](#)]
177. Hupel, M.; Lecointre, C.; Meudec, A.; Poupart, N.; Gall, E.A. Comparison of photoprotective responses to UV radiation in the brown seaweed *Pelvetia canaliculata* and the marine angiosperm *Salicornia ramosissima*. *J. Exp. Mar. Biol. Ecol.* **2011**, *401*, 36–47. [[CrossRef](#)]
178. El-Saharty, A.; Farghaly, O.A.; Hamed, A.R.; Noreldeen, H.A. Anticancer Activity of Some Marine Macroalgae in Hepatocellular Carcinoma Cell Lines (HepG2). *Int. J. Ecotoxicol. Ecobiol.* **2018**, *3*, 22–30.
179. Singh, A.; Singh, S.P.; Bamezai, R. Inhibitory potential of *Chlorella vulgaris* (E-25) on mouse skin papillomagenesis and xenobiotic detoxication system. *Anticancer Res.* **1999**, *19*, 1887–1891.
180. Mercurio, D.; Wagemaker, T.; Alves, V.; Benevenuto, C.; Gaspar, L.; Campos, P.M. In vivo photoprotective effects of cosmetic formulations containing UV filters, vitamins, *Ginkgo biloba* and red algae extracts. *J. Photochem. Photobiol. B Biol.* **2015**, *153*, 121–126. [[CrossRef](#)] [[PubMed](#)]
181. Heo, S.J.; Lee, G.W.; Song, C.B.; Jeon, Y.J. Antioxidant Activity of Enzymatic Extracts from Brown Seaweeds. *Algae* **2003**, *18*, 71–81. [[CrossRef](#)]
182. Bin, B.-H.; Kim, S.T.; Bhin, J.; Lee, T.R.; Cho, E.-G. The Development of Sugar-Based Anti-Melanogenic Agents. *Int. J. Mol. Sci.* **2016**, *17*, 583. [[CrossRef](#)]
183. Ahn, M.-J.; Yoon, K.-D.; Min, S.-Y.; Lee, J.S.; Kim, J.H.; Kim, T.G.; Kim, S.H.; Kim, N.-G.; Huh, H.; Kim, J. Inhibition of HIV-1 Reverse Transcriptase and Protease by Phlorotannins from the Brown Alga *Ecklonia cava*. *Biol. Pharm. Bull.* **2004**, *27*, 544–547. [[CrossRef](#)]
184. Xhaufilaire-Uhoda, E.; Fontaine, K.; Piérard, G. Kinetics of moisturizing and firming effects of cosmetic formulations. *Int. J. Cosmet. Sci.* **2008**, *30*, 131–138. [[CrossRef](#)] [[PubMed](#)]
185. Freile-Pelegrin, Y.; Robledo, D. Bioactive Phenolic Compounds from Algae. *Bioact. Compd. Mar. Food* **2013**, 113–129. [[CrossRef](#)]
186. Chan, Y.; Kim, K.H.; Cheah, S. Inhibitory effects of *Sargassum polycystum* on tyrosinase activity and melanin formation in B16F10 murine melanoma cells. *J. Ethnopharmacol.* **2011**, *137*, 1183–1188. [[CrossRef](#)]
187. Wang, R.; Paul, V.J.; Luesch, H. Seaweed extracts and unsaturated fatty acid constituents from the green alga *Ulva lactuca* as activators of the cytoprotective Nrf2–ARE pathway. *Free Radic. Biol. Med.* **2013**, *57*, 141–153. [[CrossRef](#)]
188. Galland-Irmouli, A.-V.; Fleurence, J.; Lamghari, R.; Luçon, M.; Rouxel, C.; Barbaroux, O.; Bronowicki, J.-P.; Villaume, C.; Guéant, J.-L. Nutritional value of proteins from edible seaweed *Palmaria palmata* (dulse). *J. Nutr. Biochem.* **1999**, *10*, 353–359. [[CrossRef](#)]
189. Samarakoon, K.; Jeon, Y.-J. Bio-functionalities of proteins derived from marine algae—A review. *Food Res. Int.* **2012**, *48*, 948–960. [[CrossRef](#)]
190. Houston, M.C. Nutraceuticals, Vitamins, Antioxidants, and Minerals in the Prevention and Treatment of Hypertension. *Prog. Cardiovasc. Dis.* **2005**, *47*, 396–449. [[CrossRef](#)] [[PubMed](#)]
191. Pereira, L. *Edible Seaweeds of the World*, 1st ed.; Taylor & Francis Group: Boca Raton, FL, USA, 2016; p. 453. ISBN 9781498730471.
192. Martínez-Hernández, G.B.; Castillejo, N.; Carrión-Monteagudo, M.D.M.; Artés, F.; Artés-Hernández, F. Nutritional and bioactive compounds of commercialized algae powders used as food supplements. *Food Sci. Technol. Int.* **2017**, *24*, 172–182. [[CrossRef](#)] [[PubMed](#)]
193. Reef, R.; Kaniewska, P.; Hoegh-Guldberg, O. Coral Skeletons Defend against Ultraviolet Radiation. *PLoS ONE* **2009**, *4*, e7995. [[CrossRef](#)] [[PubMed](#)]
194. Pereira, L. Seaweed Flora of the European North Atlantic and Mediterranean. In *Springer Handbook of Marine Biotechnology*; Se-Kwon, K., Ed.; Springer: Berlin/Heidelberg, Germany, 2015; Chapter 6; pp. 65–178. ISBN 978-3-642-53971-8.
195. Conde, F.R.; Churio, M.S.; Previtali, C.M. The photoprotector mechanism of mycosporine-like amino acids. Excited-state properties and photostability of porphyra-334 in aqueous solution. *J. Photochem. Photobiol.* **2000**, *56*, 139–144. [[CrossRef](#)]
196. De la Coba, F.; Aguilera, J.; De Galvez, M.V.; Alvarez, M.; Gallego, E.; Figueroa, F.L.; Herrera, E. Prevention of the ultraviolet effects on clinical and histopathological changes, as well as the heat shock protein-70 expression in mouse skin by topical application of algal UV-absorbing compounds. *J. Dermatol. Sci.* **2009**, *55*, 161–169. [[CrossRef](#)]
197. Jerez-Martel, I.; García-Poza, S.; Rodríguez-Martel, G.; Rico, M.; Afonso-Olivares, C.; Gómez-Pinchetti, J.L. Phenolic profile and antioxidant activity of crude extracts from microalgae and cyanobacteria strains. *J. Food Qual.* **2017**, *2017*, 2924508. [[CrossRef](#)]
198. Zolotareva, E.K.; Mokrosnop, V.M.; Stepanov, S.S. Polyphenol Compounds of Macroscopic and Microscopic Algae. *Int. J. Algae* **2019**, *21*, 5–24. [[CrossRef](#)]

199. Jean-Gilles, D.; Li, L.; Vaidyanathan, V.G.; King, R.; Cho, B.; Worthen, D.R.; Chichester, I.I.I.C.O.; Seeram, N.P. Inhibitory effects of polyphenol punicalagin on type-II collagen degradation in vitro and inflammation in vivo. *Chem.-Biol. Int.* **2013**, *25*, 90–99. [[CrossRef](#)]
200. Ryu, B.; Qian, Z.-J.; Kim, M.-M.; Nam, K.W.; Kim, S.-K. Anti-photoaging activity and inhibition of matrix metalloproteinase (MMP) by marine red alga, *Corallina pilulifera* methanol extract. *Radiat. Phys. Chem.* **2009**, *78*, 98–105. [[CrossRef](#)]
201. Kim, J.-A.; Ahn, B.-N.; Kong, C.-S.; Kim, S.-K. The chromene sargachromanol E inhibits ultraviolet A-induced ageing of skin in human dermal fibroblasts. *Br. J. Dermatol.* **2012**, *168*, 968–976. [[CrossRef](#)] [[PubMed](#)]
202. Bhatia, S.; Sharma, K.; Namdeo, A.G.; Chaugule, B.B.; Kavale, M.; Nanda, S. Broad-spectrum sun-protective action of Porphyra-334 derived from *Porphyra vietnamensis*. *Pharmacogn. Res.* **2010**, *2*, 45–49. [[CrossRef](#)] [[PubMed](#)]
203. Park, H.M.; Moon, E.; Kim, A.J.; Kim, M.H.; Lee, S.; Lee, J.B.; Park, Y.K.; Jung, H.S.; Kim, Y.B.; Kim, S.Y. Extract of *Punica granatum* inhibits skin photoaging induced by UVB irradiation. *Int. J. Dermatol.* **2010**, *49*, 276–282. [[CrossRef](#)] [[PubMed](#)]
204. Chiang, H.M.; Lin, T.J.; Chiu, C.Y.; Chang, C.W.; Hsu, K.C.; Fan, P.C.; Wen, K.C. *Coffea arabica* extract and its constituents prevent photoaging by suppressing MMPs expression and MAP kinase pathway. *Food Chem. Toxicol.* **2011**, *49*, 309–318. [[CrossRef](#)]
205. Shen, C.-T.; Chen, P.-Y.; Wu, J.-J.; Lee, T.-M.; Hsu, S.-L.; Chang, C.-M.J.; Young, C.-C.; Shieh, C.-J. Purification of algal anti-tyrosinase zeaxanthin from *Nannochloropsis oculata* using supercritical anti-solvent precipitation. *J. Supercrit. Fluids* **2011**, *55*, 955–962. [[CrossRef](#)]
206. Pavia, H.; Brock, E. Extrinsic factors influencing phlorotannin production in the brown alga *Ascophyllum nodosum*. *Mar. Ecol. Prog. Ser.* **2000**, *193*, 285–294. [[CrossRef](#)]
207. Jormalainen, V.; Honkanen, T. Variation in natural selection for growth and phlorotannins in the brown alga *Fucus vesiculosus*. *J. Evol. Biol.* **2004**, *17*, 807–820. [[CrossRef](#)]
208. Sheih, I.C.; Wu, T.K.; Fang, T.J. Antioxidant properties of a new antioxidative peptide from algae protein waste hydrolysate in different oxidation systems. *Bioresour. Technol.* **2009**, *100*, 3419–3425. [[CrossRef](#)] [[PubMed](#)]
209. Li, Y.; Qian, Z.J.; Ryu, B.; Lee, S.H.; Kim, M.M.; Kim, S.K. Chemical components and its antioxidant properties in vitro: An edible marine brown alga, *Ecklonia cava*. *Bioorg. Med. Chem.* **2009**, *17*, 1963–1973. [[CrossRef](#)] [[PubMed](#)]
210. Ferreres, F.; Lopes, G.; Gil-Izquierdo, A.; Andrade, P.B.; Sousa, C.; Mouga, T.; Valentão, P. Phlorotannin Extracts from Fucales Characterized by HPLC-DAD-ESI-MSⁿ: Approaches to Hyaluronidase Inhibitory Capacity and Antioxidant Properties. *Mar. Drugs* **2012**, *10*, 2766–2781. [[CrossRef](#)] [[PubMed](#)]
211. Sanjeewa, K.K.A.; Kim, E.A.; Son, K.T.; Jeon, Y.J. Bioactive properties and potentials cosmeceutical applications of phlorotannins isolated from brown seaweeds: A review. *J. Photochem. Photobiol. B* **2016**, *162*, 100–105. [[CrossRef](#)] [[PubMed](#)]
212. Jang, M.-S.; Park, H.-Y.; Nam, K.-H. Whitening effects of 4-hydroxyphenethyl alcohol isolated from water boiled with *Hizikia fusiformis*. *Food Sci. Biotechnol.* **2014**, *23*, 555–560. [[CrossRef](#)]
213. Brodie, J.; Williamson, C.; Barker, G.L.; Walker, R.H.; Briscoe, A.; Yallop, M. Characterising the microbiome of *Corallina officinalis*, a dominant calcified intertidal red alga. *FEMS Microbiol. Ecol.* **2016**, *92*, fiw110. [[CrossRef](#)]
214. Stengel, D.B.; Connan, S.; Popper, Z. Algal chemodiversity and bioactivity: Sources of natural variability and implications for commercial application. *Biotechnol. Adv.* **2011**, *29*, 483–501. [[CrossRef](#)]
215. Ryu, B.; Li, Y.; Qian, Z.-J.; Kim, M.-M.; Kim, S.-K. Exhibitory effects of compounds from brown alga *Ecklonia cava* on the human osteoblasts. *J. Biotechnol.* **2008**, *136*, S588. [[CrossRef](#)]
216. Heo, S.-J.; Ko, S.-C.; Kang, S.-M.; Cha, S.-H.; Lee, S.-H.; Kang, D.-H.; Jung, W.-K.; Affan, A.; Oh, C.; Jeon, Y.-J. Inhibitory effect of diphlorethohydroxycarmalol on melanogenesis and its protective effect against UV-B radiation-induced cell damage. *Food Chem. Toxicol.* **2010**, *48*, 1355–1361. [[CrossRef](#)]
217. Handelman, G.J. The evolving role of carotenoids in human biochemistry. *Nutrition* **2001**, *17*, 818–822. [[CrossRef](#)]
218. Wang, T.; Jonsdottir, R.; Ólafsdóttir, G. Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds. *Food Chem.* **2009**, *116*, 240–248. [[CrossRef](#)]
219. Lee, J.-H.; Kim, G.-H. Evaluation of Antioxidant Activity of Marine Algae-Extracts from Korea. *J. Aquat. Food Prod. Technol.* **2013**, *24*, 227–240. [[CrossRef](#)]
220. Yoshie-Stark, Y.; Hsieh, Y.P.; Suzuki, T. Distribution of flavonoids and related compounds from seaweeds in Japan. *J.-Tokyo Univ. Fish.* **2003**, *89*, 1–6.
221. Cho, S.-H.; Kang, S.-E.; Cho, J.-Y.; Kim, A.-R.; Park, S.-M.; Hong, Y.-K.; Ahn, D.-H. The Antioxidant Properties of Brown Seaweed (*Sargassum siliquastrum*) Extracts. *J. Med. Food* **2007**, *10*, 479–485. [[CrossRef](#)] [[PubMed](#)]
222. Lola-Luz, T.; Hennequart, F.; Gaffney, M. Enhancement of phenolic and flavonoid compounds in cabbage (*Brassica oleraceae*) following application of commercial seaweed extracts of the brown seaweed, (*Ascophyllum nodosum*). *Agric. Food Sci.* **2013**, *22*, 288–295. [[CrossRef](#)]
223. Corona, G.; Ji, Y.; Anegboonlap, P.; Hotchkiss, S.; Gill, C.; Yaqoob, P.; Spencer, J.P.E.; Rowland, I. Gastrointestinal modifications and bioavailability of brown seaweed phlorotannins and effects on inflammatory markers. *Br. J. Nutr.* **2016**, *115*, 1240–1253. [[CrossRef](#)] [[PubMed](#)]
224. Yan, X.; Yang, C.; Lin, G.; Chen, Y.; Miao, S.; Liu, B.; Zhao, C. Antidiabetic Potential of Green Seaweed *Enteromorpha prolifera* Flavonoids Regulating Insulin Signaling Pathway and Gut Microbiota in Type 2 Diabetic Mice. *J. Food Sci.* **2018**, *84*, 165–173. [[CrossRef](#)]

225. Farvin, K.S.; Jacobsen, C. Phenolic compounds and antioxidant activities of selected species of seaweeds from Danish coast. *Food Chem.* **2012**, *138*, 1670–1681. [[CrossRef](#)]
226. Chakraborty, K.; Maneesh, A.; Makkar, F. Antioxidant activity of brown seaweeds. *J. Aquat. Food Prod. Technol.* **2017**, *26*, 406–419. [[CrossRef](#)]
227. Vimaladevi, S.; Mahesh, A.; Dhayanithi, B.N.; Karthikeyan, N. Mosquito larvicidal efficacy of phenolic acids of seaweed *Chaetomorpha antennina* (Bory) Kuetz. against *Aedes aegypti*. *Biologia* **2012**, *67*, 212–216. [[CrossRef](#)]
228. Corsetto, P.A.; Montorfano, G.; Zava, S.; Colombo, I.; Ingadottir, B.; Jonsdottir, R.; Sveinsdottir, K.; Rizzo, A.M. Characterization of Antioxidant Potential of Seaweed Extracts for Enrichment of Convenience Food. *Antioxidants* **2020**, *9*, 249. [[CrossRef](#)] [[PubMed](#)]
229. Yoshie, Y.; Wang, W.; Hsieh, Y.P.; Suzuki, T. Compositional difference of phenolic compounds between two seaweeds, *Halimeda* spp. *J.-Tokyo Univ. Fish.* **2002**, *88*, 21–24.
230. Ilknur, A.; Turker, G. Antioxidant Activity of Five Seaweed Extracts. *New Knowl. J. Sci./Novo Znanie* **2018**, *7*, 149–155.
231. Mancini-Filho, J.; Novoa, A.V.; González, A.E.B.; de Andrade-Wartha, E.R.S.; Mancini, D.A.P. Free Phenolic Acids from the Seaweed *Halimeda monile* with Antioxidant Effect Protecting against Liver Injury. *Z. Nat. C* **2009**, *64*, 657–663. [[CrossRef](#)]
232. Keyrouz, R.; Abasq, M.L.; Le Bourvellec, C.; Blanc, N.; Audibert, L.; ArGall, E.; Hauchard, D. Total phenolic contents, radical scavenging and cyclic voltammetry of seaweeds from Brittany. *Food Chem.* **2011**, *126*, 831–836. [[CrossRef](#)]
233. Sathya, R.; Kanaga, N.; Sankar, P.; Jeeva, S. Antioxidant properties of phlorotannins from brown seaweed *Cystoseira trinodis* (Forsskål) C. Agardh. *Arab. J. Chem.* **2017**, *10*, S2608–S2614. [[CrossRef](#)]
234. Yoon, N.Y.; Chung, H.Y.; Kim, H.R.; Choi, J.S. Acetyl- and butyrylcholinesterase inhibitory activities of sterols and phlorotannins from *Ecklonia stolonifera*. *Fish. Sci.* **2008**, *74*, 200–207. [[CrossRef](#)]
235. Connan, S.; Goulard, F.; Stiger, V.; Deslandes, E.; Gall, E.A. Interspecific and temporal variation in phlorotannin levels in an assemblage of brown algae. *Bot. Mar.* **2004**, *47*, 410–416. [[CrossRef](#)]
236. Horincar, V.B.; Parfene, G.; Bahrim, G. Evaluation of bioactive compounds in extracts obtained from three Romanian marine algae species. *Rom. Biotechnol. Lett.* **2011**, *16*, 71–78.
237. Watson, D.C.; Norton, T.A. Dietary preferences of the common periwinkle, *Littorina littorea* (L.). *J. Exp. Mar. Biol. Ecol.* **1985**, *88*, 193–211. [[CrossRef](#)]
238. Li, Y.; Fu, X.; Duan, D.; Liu, X.; Xu, J.; Gao, X. Extraction and Identification of Phlorotannins from the Brown Alga, *Sargassum fusiforme* (Harvey) Setchell. *Mar. Drugs* **2017**, *15*, 49. [[CrossRef](#)]
239. Kim, K.-N.; Yang, H.-M.; Kang, S.-M.; Kim, D.; Ahn, G.; Jeon, Y.-J. Octaphlorethol A isolated from *Ishige foliacea* inhibits α -MSH-stimulated induced melanogenesis via ERK pathway in B16F10 melanoma cells. *Food Chem. Toxicol.* **2013**, *59*, 521–526. [[CrossRef](#)] [[PubMed](#)]
240. Kim, J.S.; Lee, J.H. Antioxidant and anti-inflammatory activity of phloroglucinol from seaweeds. *FASEB J.* **2016**, *30*, 1174.12.
241. Samri, N.; Hsaine, L.; El Kafhi, S.; Khelifi, S.; Etahiri, S. Radical Scavenging Activity and Phenolic Contents of Brown Seaweeds Harvested from the Coast of Sidi Bouzid (El Jadida, Morocco). *Int. J. Pharm. Sci. Rev. Res.* **2019**, *54*, 116–122.
242. Zubia, M.; Payri, C.; Deslandes, E. Alginate, mannitol, phenolic compounds and biological activities of two range-extending brown algae, *Sargassum mangarevense* and *Turbinaria ornata* (Phaeophyta: Fucales), from Tahiti (French Polynesia). *Environ. Boil. Fishes* **2008**, *20*, 1033–1043. [[CrossRef](#)]
243. Surget, G.; Roberto, V.; Le Lann, K.; Mira, S.; Guérard, F.; Laizé, V.; Poupart, N.; Cancela, M.L.; Stiger-Pouvreau, V. Marine green macroalgae: A source of natural compounds with mineralogenic and antioxidant activities. *Environ. Boil. Fishes* **2016**, *29*, 575–584. [[CrossRef](#)]
244. Heo, S.-J.; Ko, S.-C.; Kang, S.-M.; Kang, H.-S.; Kim, J.-P.; Kim, S.-H.; Lee, K.-W.; Cho, M.-G.; Jeon, Y.-J. Cytoprotective effect of fucoxanthin isolated from brown algae *Sargassum siliquastrum* against H₂O₂-induced cell damage. *Eur. Food Res. Technol.* **2008**, *228*, 145–151. [[CrossRef](#)]
245. Miranda, A.; Montoya, M.; Araos, M.; Mellado, M.; Villena, J.; Arancibia, P.; Madrid, A.; Jara, C. Antioxidant and anti cancer activities of brown and red seaweed extracts from Chilean coasts. *Lat. Am. J. Aquat. Res.* **2018**, *46*, 301–313. [[CrossRef](#)]
246. Abdala-Díaz, R.T.; Cabellopasini, A.; Perezrodriguez, E.; Álvarez, R.M.C.; Figueroa, F.D.L. Daily and seasonal variations of optimum quantum yield and phenolic compounds in *Cystoseira tamariscifolia* (Phaeophyta). *Mar. Biol.* **2005**, *148*, 459–465. [[CrossRef](#)]
247. Sugiura, Y.; Tanaka, R.; Katsuzaki, H.; Imai, K.; Matsushita, T. The anti-inflammatory effects of phlorotannins from *Eisenia arborea* on mouse ear edema by inflammatory inducers. *J. Funct. Foods* **2013**, *5*, 2019–2023. [[CrossRef](#)]
248. Cian, R.E.; Caballero, M.S.; Sabbag, N.; González, R.J.; Drago, S.R. Bio-accessibility of bioactive compounds (ACE inhibitors and antioxidants) from extruded maize products added with a red seaweed *Porphyra columbina*. *LWT-Food Sci. Technol.* **2014**, *55*, 51–58. [[CrossRef](#)]
249. Christaki, E.; Bonos, E.; Giannenas, I.; Florou-Paneri, P. Functional properties of carotenoids originating from algae. *J. Sci. Food Agric.* **2012**, *93*, 5–11. [[CrossRef](#)] [[PubMed](#)]
250. Pallela, R.; Na-Young, Y.; Kim, S.-K. Anti-photoaging and Photoprotective Compounds Derived from Marine Organisms. *Mar. Drugs* **2010**, *8*, 1189–1202. [[CrossRef](#)]
251. Stolz, P.; Obermayer, B. Manufacturing microalgae for skincare. *Cosmet. Toilet.* **2005**, *120*, 99–106.
252. Wang, H.-M.D.; Chen, C.-C.; Huynh, P.; Chang, J.-S. Exploring the potential of using algae in cosmetics. *Bioresour. Technol.* **2014**, *184*, 355–362. [[CrossRef](#)]

253. Alam, T.; Najam, L.; Al-Harrasi, A. Extraction of natural pigments from marine algae. *J. Agric. Mar. Sci.* **2018**, *23*, 81–91. [CrossRef]
254. Ji, N.K.; Kumar, R.N.; Bora, A.; Amb, M.K.; Chakraborty, S. An Evaluation of the Pigment Composition of Eighteen Marine Macroalgae Collected from Okha Coast, Gulf of Kutch, India. *Our Nat.* **1970**, *7*, 48–55. [CrossRef]
255. Borowitzka, M. Microalgae as sources of pharmaceuticals and other biologically active compounds. *Environ. Boil. Fishes* **1995**, *7*, 3–15. [CrossRef]
256. Metting, B.; Pyne, J.W. Biologically active compounds from microalgae. *Enzym. Microb. Technol.* **1986**, *8*, 386–394. [CrossRef]
257. Cannell, R.J.P. Algae as a source of biologically active products. *Pestic. Sci.* **1993**, *39*, 147–153. [CrossRef]
258. Paul, C.; Pohnert, G. Production and role of volatile halogenated compounds from marine algae. *Nat. Prod. Rep.* **2011**, *28*, 186–195. [CrossRef] [PubMed]
259. Ibañez, E.; Herrero, M.; Mendiola, J.A.; Castro-Puyana, M. Extraction and Characterization of Bioactive Compounds with Health Benefits from Marine Resources: Macro and Micro Algae, Cyanobacteria, and Invertebrates. In *Marine Bioactive Compounds*; Springer: Boston, MA, USA, 2011; pp. 55–98. [CrossRef]
260. Vo, T.-S.; Ngo, D.-H.; Kim, S.-K. Marine algae as a potential pharmaceutical source for anti-allergic therapeutics. *Process Biochem.* **2012**, *47*, 386–394. [CrossRef]
261. Osório, C.; Machado, S.; Peixoto, J.; Bessada, S.; Pimentel, F.B.; Alves, R.C.; Oliveira, M.B.P.P. Pigments Content (Chlorophylls, Fucoxanthin and Phycobiliproteins) of Different Commercial Dried Algae. *Separations* **2020**, *7*, 33. [CrossRef]
262. Go, H.; Hwang, H.-J.; Nam, T.-J. A glycoprotein from *Laminaria japonica* induces apoptosis in HT-29 colon cancer cells. *Toxicol. Vitro.* **2010**, *24*, 1546–1553. [CrossRef]
263. Ermakova, S.; Sokolova, R.; Kim, S.-M.; Um, B.-H.; Isakov, V.; Zvyagintseva, T. Fucoindans from Brown Seaweeds *Sargassum hornery*, *Eclonia cava*, *Costaria costata*: Structural Characteristics and Anticancer Activity. *Appl. Biochem. Biotechnol.* **2011**, *164*, 841–850. [CrossRef]
264. Costa, L.S.; Fidelis, G.P.; Telles, C.B.; Dantas-Santos, N.; Camara, R.B.; Cordeiro, S.L.; Costa, M.S.; Almeida-Lima, J.; Melo-Silveira, R.F.; Oliveira, R.M.; et al. Antioxidant and antiproliferative activities of heterofucans from the seaweed *Sargassum filipendula*. *Mar. Drugs* **2011**, *9*, 952–966. [CrossRef]
265. Satomi, Y. Fucoxanthin induces GADD45A expression and G1 arrest with SAPK/JNK activation in LNCap human prostate cancer cells. *Anticancer Res.* **2012**, *32*, 807–813. [PubMed]
266. Kim, J.-Y.; Yoon, M.-Y.; Cha, M.-R.; Hwang, J.-H.; Park, E.; Choi, S.-U.; Park, H.-R.; Hwang, Y.-I. Methanolic Extracts of *Plocamium telfairiae* Induce Cytotoxicity and Caspase-Dependent Apoptosis in HT-29 Human Colon Carcinoma Cells. *J. Med. Food* **2007**, *10*, 587–593. [CrossRef]
267. Takaichi, S. Distributions, biosyntheses, and functions of carotenoids in algae. *Agro Food Ind. Hi-Tech* **2013**, *24*, 55–58. [CrossRef]
268. Quilodrán, B.; Hinzpeter, I.; Hormazabal, E.; Quiroz, A.; Shene, C. Docosahexaenoic acid (C22: 6n–3, DHA) and astaxanthin production by *Thraustochytriidae* sp. AS4-A1 a native strain with high similitude to *Ulkenia* sp.: Evaluation of liquid residues from the food industry as nutrient sources. *Enzym. Microb. Technol.* **2010**, *47*, 24–30. [CrossRef]
269. Amon, J.P.; French, K.H. Photoresponses of the Marine Protist *Ulkenia* sp. Zoospores to Ambient, Artificial and Bioluminescent Light. *Mycologia* **2004**, *96*, 463. [CrossRef]
270. Hosikian, A.; Lim, S.; Halim, R.; Danquah, M. Chlorophyll Extraction from Microalgae: A Review on the Process Engineering Aspects. *Int. J. Chem. Eng.* **2010**, *2010*, 391632. [CrossRef]
271. Spears, K. Developments in food colorings: The natural alternatives. *Trends Biotechnol.* **1988**, *6*, 283–288. [CrossRef]
272. La-Mer. My Skin—And What It Needs. 2018. Available online: <https://www.la-mer.com/en> (accessed on 22 September 2018).
273. Lanfer-Marquez, U.M.; Barros, R.M.; Sinnecker, P. Antioxidant activity of chlorophylls and their derivatives. *Food Res. Int.* **2005**, *38*, 885–891. [CrossRef]
274. Horwitz, B. Role of chlorophyll in proctology. *Am. J. Surg.* **1951**, *81*, 81–84. [CrossRef]
275. Kawata, A.; Murakami, Y.; Suzuki, S.; Fujisawa, S. Anti-inflammatory activity of β -carotene, lycopene and tri-n-butylborane, a scavenger of reactive oxygen species. *In Vivo* **2018**, *32*, 255–264.
276. Borowitzka, M.A. High-value products from microalgae—Their development and commercialization. *J. Appl. Phycol.* **2013**, *25*, 743–756. [CrossRef]
277. Sies, H.; Stahl, W. Carotenoids and UV protection. *Photochem. Photobiol. Sci.* **2004**, *3*, 749–752. [CrossRef] [PubMed]
278. Spolaore, P.; Joannis-Cassan, C.; Duran, E.; Isambert, A. Commercial applications of microalgae. *J. Biosci. Bioeng.* **2006**, *101*, 87–96. [CrossRef]
279. Peng, J.; Yuan, J.-P.; Wu, C.-F.; Wang, J.-H. Fucoxanthin, a marine carotenoid present in brown seaweeds and diatoms: Me-tabolism and bioactivities relevant to human health. *Mar. Drugs* **2011**, *9*, 1806–1828. [CrossRef]
280. D’Orazio, N.; Gemello, E.; Gammone, M.A.; De Girolamo, M.; Ficoneri, C.; Riccioni, G. Fucoxanthin: A Treasure from the Sea. *Mar. Drugs* **2012**, *10*, 604–616. [CrossRef] [PubMed]
281. Kirti, K.; Amita, S.; Priti, S.; Kumar, A.M.; Jyoti, S. Colorful World of Microbes: Carotenoids and Their Applications. *Adv. Biol.* **2014**, *2014*, 837891. [CrossRef]
282. Morabito, K.; Shapley, N.C.; Steeley, K.G.; Tripathi, A. Review of sunscreen and the emergence of non-conventional absorbers and their applications in ultraviolet protection. *Int. J. Cosmet. Sci.* **2011**, *33*, 385–390. [CrossRef] [PubMed]
283. Chinnadurai, S.; Kalyanasundaram, G. Estimation of major pigment content in seaweeds collected from Pondicherry coast. *Int. J. Sci. Technol.* **2013**, *9*, 522–525.

284. Von, E.; McDowell, R.H. *Chemistry and Enzymology of Marine Algal Polysaccharides*; Academic Press: London, UK; New York, NY, USA, 1967.
285. Ponce, N.M.; Pujol, C.A.; Damonte, E.B.; Flores, M.L.; Stortz, C.A. Fucoidans from the brown seaweed *Adenocystis utricularis*: Extraction methods, antiviral activity and structural studies. *Carbohydr. Res.* **2003**, *338*, 153–165. [[CrossRef](#)]
286. Jayasankar, R.; Ramalingam, J.R. Photosynthetic pigment of marine algae from Mandapam coast. *Seaweed Res. Util.* **1993**, *16*, 41–43.
287. Sudhakar, M.P.; Ananthalakshmi, J.S.; Nair, B.B. Extraction, purification, and study on antioxidant properties of fucoxanthin from brown seaweeds. *J. Chem. Pharm. Res.* **2013**, *5*, 169–175.
288. Panjaitan, R.S. Pigment contents of *Sargassum polycistum* macroalgae lipid from Sayang heulang beach, Indonesia. *Sci. Study Res. Chem. Chem. Eng. Biotechnol. Food Ind.* **2019**, *20*, 365–375.
289. O'Connor, I.; O'Brien, N. Modulation of UVA light-induced oxidative stress by β -carotene, lutein and astaxanthin in cultured fibroblasts. *J. Dermatol. Sci.* **1998**, *16*, 226–230. [[CrossRef](#)]
290. Gevaert, F.; Creach, A.; Davoult, D.; Holl, A.C.; Seuront, L.; Lemoine, Y. Photo-inhibition and seasonal photosynthetic performance of the seaweed *Laminaria saccharina* during a simulated tidal cycle: Chlorophyll fluorescence measurements and pigment analysis. *Plant Cell Environ.* **2002**, *25*, 859–872. [[CrossRef](#)]
291. Pessoa, M.F. Harmful effects of UV radiation in algae and aquatic macrophytes—A review. *Emir. J. Food Agric.* **2012**, *24*, 510–526. [[CrossRef](#)]
292. Indriatmoko, M.A.; Indrawati, R.; Limantara, L. Composition of the Main Dominant Pigments from Potential Two Edible Sea-weeds. *Philipp. J. Sci.* **2018**, *147*, 47–55.

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